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(71)	Applicant(s)  Immunex Corporation
(72)	Inventor(s)  Dirk M. Anderson; Laurent J. Galibert; Eugene Maraskovsky
(74)	Agent/Attorney PHILLIPS ORMONDE and FITZPATRICK, 367 Collins Street, MRLBOURNE VIC 3000



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(71) Applicant: IMMUNEX COF 51 University Street, Seat		aw Dep	Published With international search report.	
(72) Inventors: ANDERSON, I Seattle, WA 98107 (US). Avenue West, Seattle, W. Eugene; 4123 Evanston (US).	GALIBERT, Laurent, J. 98119 (US). MARASK	h /.	fional search report: 1 October 1998 (01.10.98)	
(74) Agent: PERKINS, Patricia, Dept., 51 University Street			w	·

(54) Title: LIGAND FOR RECEPTOR ACTIVATOR OF NF-KAPPA B, LIGAND IS MEMBER OF THE SUPERFAMILY

#### (57) Abstrac

Isolated ligands, DNAs encoding such ligands, and pharmaceutical compositions made therefrom, are disclosed. The isolated ligands can be used to regulate an immune response. The ligands are also useful in screening for inhibitors thereof.

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#### TITLE

LIGAND FOR RECEPTOR ACTIVATOR OF NF-KAPPA B, LIGAND IS MEMBER OF THE SUPERFAMILY

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# TECHNICAL FIELD OF THE INVENTION

The present invention relates generally to the field of cytokines, and more specifically to cytokine receptor/ligand pairs having immunoregulatory activity.

# BACKGROUND OF THE INVENTION

Efficient functioning of the immune system requires a fine balance between cell proliferation and differentiation and cell death, to ensure that the immune system is capable of reacting to foreign, but not self antigens. Integral to the process of regulating the immune and inflammatory response are various members of the Tumor Necrosis Factor (TNF) Receptor/Nerve Growth Factor Receptor superfamily (Smith et al., Science 248:1019; 1990). This family of receptors includes two different TNF receptors (Type I and Type II; Smith et al., supra; and Schall et al., Cell 61:361, 1990), nerve growth factor receptor (Johnson et al., Cell 47:545, 1986), B cell antigen CD40 (Stamenkovic et al., EMBO J. 8:1403, 1989), CD27 (Camerini et al., J. Immunol. 147:3165, 1991), CD30 (Durkop et al., Cell 68:421, 1992), T cell antigen OX40 (Mallett et al., EMBO J. 9:1063, 1990), human Fas antigen (Itoh et al., Cell 66:233, 1991), murine 4-1BB receptor (Kwon et al., Proc. Natl. Acad. Sci. USA 86:1963, 1989) and a receptor referred to as Apoptosis-Inducing Receptor (AIR; USSN 08/720,864, filed October 4, 1996).

CD40 is a receptor present on B lymphocytes, epithelial cells and some carcinoma cell lines that interacts with a ligand found on activated T cells, CD40L (USSN 08/249,189, filed May 24, 1994). The interaction of this ligand/receptor pair is essential for both the cellular and humoral immune response. Signal transduction via CD40 is mediated through the association of the cytoplasmic domain of this molecule with members of the TNF receptor-associated factors (TRAFs; Baker and Reddy, Oncogene 12:1, 1996). It has recently been found that mice that are defective in TRAF3 expression due to a targeted disruption in the gene encoding TRAF3 appear normal at birth but develop progressive hypoglycemia and depletion of peripheral white cells, and die by about ten days of age (Xu et al., Immunity 5:407, 1996). The immune responses of chimeric mice reconstituted with TRAF3-/- fetal liver cells resemble those of CD40-deficient mice, although TRAF3-/- B cells appear to be functionally normal.

The critical role of TRAF3 in signal transduction may be in its interaction with one of the other members of the TNF receptor superfamily, for example, CD30 or CD27,

which are present on T cells. Alternatively, there may be other, as yet unidentified members of this family of receptors that interact with TRAF3 and play an important role in postnatal development as well as in the development of a competent immune system. Identifying additional members of the TNF receptor superfamily would provide an additional means of regulating the immune and inflammatory response, as well as potentially providing further insight into post-natal development in mammals.

# SUMMARY OF THE INVENTION

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The present invention provides a counterstructure, or ligand, for a novel receptor referred to as RANK (for receptor activator of NF-kB), that is a member of the TNF superfamily. The ligand, which is referred to as RANKL, is a Type 2 transmembrane protein with an intracellular domain of less than about 50 amino acids, a transmembrane domain and an extracellular domain of from about 240 to 250 amino acids. Similar to other members of the TNF family to which it belongs, RANKL has a 'spacer' region between the transmembrane domain and the receptor binding domain that is not necessary for receptor binding. Accordingly, soluble forms of RANKL can comprise the entire extracellular domain or fragments thereof that include the receptor binding region.

RANK is a Type I transmembrane protein having 616 amino acid residues that is a member of the TNFR superfamily, and interacts with TRAF3. Triggering of RANK by over-expression, co-expression of RANK and membrane bound RANKL, or by soluble RANKL or agonistic antibodies to RANK, results in the upregulation of the transcription factor NF-kB, a ubiquitous transcription factor that is most extensively utilized in cells of the immune system.

These and other aspects of the present invention will become evident upon reference to the following detailed description of the invention.

# BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 demonstrates the influence of RANK.Fc and hRANKL on activated T cell growth. Human peripheral blood T cells were cultured as described in Example 12; viable T cell recovery was determined by triplicate trypan blue countings.

Figure 2 illustrates the ability of RANKL to induce human DC cluster formation. Functionally mature dendritic cells (DC) were generated *in vitro* from CD34<sup>+</sup> bone marrow (BM) progenitors and cultured as described in Example 13. CD1a<sup>+</sup> DC were cultured in a cytokine cocktail alone (Figure 2A), in cocktail plus CD40L (Figure 2B), RANKL (Figure 2C), or heat inactivated (ΔH) RANKL (Figure 2D), and then photographed using an inversion microscope.

Figure 3 demonstrates that RANKL enhances DC allo-stimulatory capacity. Allogeneic T cells were incubated with varying numbers of irradiated DC cultured as

described in Example 13. The cultures were pulsed with [3H]-thyrnidine and the cells harvested onto glass fiber sheets for counting. Values represent the mean  $\pm$  standard deviation (SD) of triplicate cultures.

Figure 4 presents an alignment of human RANK with other TNFR family members in the region of structurally conserved extracellular cysteine-rich pseudorepeats. Predicted disulfide linkages (DS1-DS3) are indicated. RANK and CD40 contain identical amino acid substitutions (C^H, C^G) eliminating DS2 in the second pseudorepeat.

Figure 5 presents an alignment of human RANKL with other TNF family members.

#### DETAILED DESCRIPTION OF THE INVENTION

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A novel partial cDNA insert with a predicted open reading frame having some similarity to CD40 was identified in a database containing sequence information from cDNAs generated from human bone marrow-derived dendritic cells (DC). The insert was used to hybridize to colony blots generated from a DC cDNA library containing full-length cDNAs. Several colony hybridizations were performed, and two clones (SEQ ID NOs:1 and 3) were isolated. SEQ ID NO:5 shows the nucleotide and amino acid sequence of a predicted full-length protein based on alignment of the overlapping sequences of SEQ ID NOs:1 and 3.

RANK is a member of the TNF receptor superfamily; it most closely resembles CD40 in the extracellular region. Similar to CD40, RANK associates with TRAF2 and TRAF3 (as determined by co-immunoprecipitation assays substantially as described by Rothe et al., Cell 83:1243, 1995). TRAFs are critically important in the regulation of the immune and inflammatory response. Through their association with various members of the TNF receptor superfamily, a signal is transduced to a cell. That signal results in the proliferation, differentiation or apoptosis of the cell, depending on which receptor(s) is/are triggered and which TRAF(s) associate with the receptor(s); different signals can be transduced to a cell via coordination of various signaling events. Thus, a signal transduced through one member of this family may be proliferative, differentiative or apoptotic, depending on other signals being transduced to the cell, and/or the state of differentiation of the cell. Such exquisite regulation of this proliferative/apoptotic pathway is necessary to develop and maintain protection against pathogens; imbalances can result in autoimmune disease.

RANK is expressed on epithelial cells, some B cell lines, and on activated T cells. However, its expression on activated T cells is late, about four days after activation. This time course of expression coincides with the expression of Fas, a known agent of apoptosis. RANK may act as an anti-apoptotic signal, rescuing cells that express RANK from apoptosis as CD40 is known to do. Alternatively, RANK may confirm an apoptotic

signal under the appropriate circumstances, again similar to CD40. RANK and its ligand are likely to play an integral role in regulation of the immune and inflammatory response.

Moreover, the post-natal lethality of mice having a targeted disruption of the TRAF3 gene demonstrates the importance of this molecule not only in the immune response but in development. The isolation of RANK, as a protein that associates with TRAF3, and its ligand, RANKL, will allow further definition of this signaling pathway, and development of diagnostic and therapeutic modalities for use in the area of autoimmune and/or inflammatory disease.

#### 10 DNAs, Proteins and Analogs

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The present invention provides isolated RANKL polypeptides and analogs (or muteins) thereof having an activity exhibited by the native molecule (i.e., RANKL muteins that bind specifically to a RANK expressed on cells or immobilized on a surface or to RANKL-specific antibodies; soluble forms thereof that inhibit RANK ligand-induced signaling through RANK). Such proteins are substantially free of contaminating endogenous materials and, optionally, without associated native-pattern glycosylation. Derivatives of RANKL within the scope of the invention also include various structural forms of the primary proteins which retain biological activity. Due to the presence of ionizable amino and carboxyl groups, for example, a RANKL protein may be in the form of acidic or basic salts, or may be in neutral form. Individual amino acid residues may also be modified by oxidation or reduction. The primary amino acid structure may be modified by forming covalent or aggregative conjugates with other chemical moieties, such as glycosyl groups, lipids, phosphate, acetyl groups and the like, or by creating amino acid sequence mutants. Covalent derivatives are prepared by linking particular functional groups to amino acid side chains or at the N- or C-termini.

Derivatives of RANKL may also be obtained by the action of cross-linking agents, such as M-maleimidobenzoyl succinimide ester and N-hydroxysuccinimide, at cysteine and lysine residues. The inventive proteins may also be covalently bound through reactive side groups to various insoluble substrates, such as cyanogen bromide-activated, bisoxirane-activated, carbonyldiimidazole-activated or tosyl-activated agarose structures, or by adsorbing to polyolefin surfaces (with or without glutaraldehyde cross-linking). Once bound to a substrate, the proteins may be used to selectively bind (for purposes of assay or purification) antibodies raised against the proteins or against other proteins which are similar to RANKL, as well as other proteins that bind RANKL or homologs thereof.

Soluble forms of RANKL are also within the scope of the invention. The nucleotide and predicted amino acid sequence of the RANKL is shown in SEQ ID NOs:10 and 12 (murine and human, respectively). Computer analysis indicated that the RANKL is a Type 2 transmembrane protein; murine RANKL contains a predicted 48 amino acid

intracellular domain, 21 amino acid transmembrane domain and 247 amino acid extracellular domain, and human RANKL contains a predicted 47 amino acid intracellular domain, 21 amino acid transmembrane domain and 249 amino acid extracellular domain.

Soluble RANKL comprises a signal peptide and the extracellular domain or a fragment thereof. An exemplary signal peptide is that shown in SEQ ID NO:9; other signal (or leader) peptides are well-known in the art, and include that of murine Interleukin-7 or human growth hormone. RANKL is similar to other members of the TNF family in having a region of amino acids between the transmembrane domain and the receptor binding region that does not appear to be required for biological activity; this is referred to as a 'spacer' region. Amino acid sequence alignment indicates that the receptor binding region is from about amino acid 162 of human RANKL to about amino acid 317 (corresponding to amino acid 139 through 294 of murine RANKL, SEQ ID NO:10), beginning with an Ala residue that is conserved among many members of the family (amino acid 162 of SEQ ID NO:12).

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Moreover, fragments of the extracellular domain will also provide soluble forms of RANKL. Those skilled in the art will recognize that the actual receptor binding region may be different than that predicted by computer analysis. Thus, the N-terminal amino acid of a soluble RANKL is expected to be within about five amino acids on either side of the conserved Ala residue. Alternatively, all or a portion of the spacer region may be included at the N-terminus of a soluble RANKL, as may be all or a portion of the transmembrane and/or intracellular domains, provided that the resulting soluble RANKL is not membrane-associated. Accordingly, a soluble RANKL will have an N-terminal amino acid selected from the group consisting of amino acids 1 through 162 of SEQ ID NO:12 (1 though 139 of SEQ ID NO:10). Preferably, the amino terminal amino acid is between amino acids 69 and 162 of SEQ ID NO:12 (human RANKL; amino acids 48 and 139 of SEQ ID NO:10). Similarly, the carboxy terminal amino acid can be between amino acid 313 and 317 of SEQ ID NO:12 (human RANKL; corresponding to amino acids 290 through 294 of SEQ ID NO:10). Those skilled in the art can prepare these and additional soluble forms through routine experimentation.

Fragments can be prepared using known techniques to isolate a desired portion of the extracellular region, and can be prepared, for example, by comparing the extracellular region with those of other members of the TNF family (of which RANKL is a member) and selecting forms similar to those prepared for other family members. Alternatively, unique restriction sites or PCR techniques that are known in the art can be used to prepare numerous truncated forms which can be expressed and analyzed for activity.

Other derivatives of the RANKL proteins within the scope of this invention include covalent or aggregative conjugates of the proteins or their fragments with other proteins or polypeptides, such as by synthesis in recombinant culture as N-terminal or C-terminal fusions. For example, the conjugated peptide may be a signal (or leader) polypeptide

sequence at the N-terminal region of the protein which co-translationally or post-translationally directs transfer of the protein from its site of synthesis to its site of function inside or outside of the cell membrane or wall (e.g., the yeast \u03c4-factor leader).

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Protein fusions can comprise peptides added to facilitate purification or identification of RANKL proteins and homologs (e.g., poly-His). The amino acid sequence of the inventive proteins can also be linked to an identification peptide such as that described by Hopp et al., *Bio/Technology* 6:1204 (1988). Such a highly antigenic peptide provides an epitope reversibly bound by a specific monoclonal antibody, enabling rapid assay and facile purification of expressed recombinant protein. The sequence of Hopp et al. is also specifically cleaved by bovine mucosal enterokinase, allowing removal of the peptide from the purified protein. Fusion proteins capped with such peptides may also be resistant to intracellular degradation in *E. coli*.

Fusion proteins further comprise the amino acid sequence of a RANKL linked to an immunoglobulin Fc region. An exemplary Fc region is a human IgG<sub>1</sub> having a nucleotide an amino acid sequence set forth in SEQ ID NO:8. Fragments of an Fc region may also be used, as can Fc muteins. For example, certain residues within the hinge region of an Fc region are critical for high affinity binding to Fc<sub>7</sub>RI. Canfield and Morrison (*J. Exp. Med.* 173:1483; 1991) reported that Leu<sub>(234)</sub> and Leu<sub>(235)</sub>were critical to high affinity binding of IgG<sub>3</sub> to Fc<sub>7</sub>RI present on U937 cells. Similar results were obtained by Lund et al. (*J. Immunol.* 147:2657, 1991; *Molecular Immunol.* 29:53, 1991). Such mutations, alone or in combination, can be made in an IgG<sub>1</sub> Fc region to decrease the affinity of IgG<sub>1</sub> for FcR. Depending on the portion of the Fc region used, a fusion protein may be expressed as a dimer, through formation of interchain disulfide bonds. If the fusion proteins are made with both heavy and light chains of an antibody, it is possible to form a protein oligomer with as many as four RANKL regions.

In another embodiment, RANKL proteins further comprise an oligomerizing peptide such as a leucine zipper domain. Leucine zippers were originally identified in several DNA-binding proteins (Landschulz et al., Science 240:1759, 1988). Leucine zipper domain is a term used to refer to a conserved peptide domain present in these (and other) proteins, which is responsible for dimerization of the proteins. The leucine zipper domain (also referred to herein as an oligomerizing, or oligomer-forming, domain) comprises a repetitive heptad repeat, with four or five leucine residues interspersed with other amino acids. Examples of leucine zipper domains are those found in the yeast transcription factor GCN4 and a heat-stable DNA-binding protein found in rat liver (C/EBP; Landschulz et al., Science 243:1681, 1989). Two nuclear transforming proteins, for and jun, also exhibit leucine zipper domains, as does the gene product of the murine proto-oncogene, c-myc (Landschulz et al., Science 240:1759, 1988). The products of the muclear oncogenes for and jun comprise leucine zipper domains preferentially form a

heterodimer (O'Shea et al., Science 245:646, 1989; Turner and Tjian, Science 243:1689, 1989). The leucine zipper domain is necessary for biological activity (DNA binding) in these proteins.

The fusogenic proteins of several different viruses, including paramyxovirus, coronavirus, measles virus and many retroviruses, also possess leucine zipper domains (Buckland and Wild, Nature 338:547,1989; Britton, Nature 353:394, 1991; Delwart and Mosialos, AIDS Research and Human Retroviruses 6:703, 1990). The leucine zipper domains in these fusogenic viral proteins are near the transmembrane region of the proteins; it has been suggested that the leucine zipper domains could contribute to the oligomeric structure of the fusogenic proteins. Oligomerization of fusogenic viral proteins is involved in fusion pore formation (Spruce et al, Proc. Natl. Acad. Sci. U.S.A. 88:3523, 1991). Leucine zipper domains have also been recently reported to play a role in oligomerization of heat-shock transcription factors (Rabindran et al., Science 259:230, 1993).

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Leucine zipper domains fold as short, parallel coiled coils. (O'Shea et al., Science 254:539; 1991) The general architecture of the parallel coiled coil has been well characterized, with a "knobs-into-holes" packing as proposed by Crick in 1953 (Acta Crystallogr. 6:689). The dimer formed by a leucine zipper domain is stabilized by the heptad repeat, designated  $(abcdefg)_n$  according to the notation of McLachlan and Stewart (J. Mol. Biol. 98:293; 1975), in which residues a and d are generally hydrophobic residues, with d being a leucine, which line up on the same face of a helix. Oppositely-charged residues commonly occur at positions g and e. Thus, in a parallel coiled coil formed from two helical leucine zipper domains, the "knobs" formed by the hydrophobic side chains of the first helix are packed into the "holes" formed between the side chains of the second helix.

The leucine residues at position d contribute large hydrophobic stabilization energies, and are important for dimer formation (Krystek et al., Int. J. Peptide Res. 38:229, 1991). Lovejoy et al. recently reported the synthesis of a triple-stranded  $\alpha$ -helical bundle in which the helices run up-up-down (Science 259:1288, 1993). Their studies confirmed that hydrophobic stabilization energy provides the main driving force for the formation of coiled coils from helical monomers. These studies also indicate that electrostatic interactions contribute to the stoichiometry and geometry of coiled coils.

Several studies have indicated that conservative amino acids may be substituted for individual leucine residues with minimal decrease in the ability to dimerize; multiple changes, however, usually result in loss of this ability (Landschulz et al., Science 243:1681, 1989; Turner and Tjian, Science 243:1689, 1989; Hu et al., Science 250:1400, 1990). van Heckeren et al. reported that a number of different amino residues can be substituted for the leucine residues in the leucine zipper domain of GCN4, and further found that some GCN4 proteins containing two leucine substitutions were weakly active

(Nucl. Acids Res. 20:3721, 1992). Mutation of the first and second heptadic leucines of the leucine zipper domain of the measles virus fusion protein (MVF) did not affect syncytium formation (a measure of virally-induced cell fusion); however, mutation of all four leucine residues prevented fusion completely (Buckland et al., J. Gen. Virol. 73:1703, 1992). None of the mutations affected the ability of MVF to form a tetramer.

Amino acid substitutions in the a and d residues of a synthetic peptide representing the GCN4 leucine zipper domain have been found to change the oligomerization properties of the leucine zipper domain (Alber, Sixth Symposium of the Protein Society, San Diego, CA). When all residues at position a are changed to isoleucine, the leucine zipper still forms a parallel dimer. When, in addition to this change, all leucine residues at position a are also changed to isoleucine, the resultant peptide spontaneously forms a trimeric parallel coiled coil in solution. Substituting all amino acids at position a with leucine results in a peptide that tetramerizes. Peptides containing these substitutions are still referred to as leucine zipper domains.

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The present invention also includes RANKL with or without associated native-pattern glycosylation. Proteins expressed in yeast or mammalian expression systems, e.g., COS-7 cells, may be similar or slightly different in molecular weight and glycosylation pattern than the native molecules, depending upon the expression system. Expression of DNAs encoding the inventive proteins in bacteria such as *E. coli* provides non-glycosylated molecules. Functional mutant analogs of RANKL protein having inactivated N-glycosylation sites can be produced by oligonucleotide synthesis and ligation or by site-specific mutagenesis techniques. These analog proteins can be produced in a homogeneous, reduced-carbohydrate form in good yield using yeast expression systems. N-glycosylation sites in eukaryotic proteins are characterized by the amino acid triplet Asn-A<sub>1</sub>-Z, where A<sub>1</sub> is any amino acid except Pro, and Z is Ser or Thr. In this sequence, asparagine provides a side chain amino group for covalent attachment of carbohydrate. Such a site can be eliminated by substituting another amino acid for Asn or for residue Z, deleting Asn or Z, or inserting a non-Z amino acid between A<sub>1</sub> and Z, or an amino acid other than Asn between Asn and A<sub>1</sub>.

RANKL protein derivatives may also be obtained by mutations of the native RANKL or subunits thereof. A RANKL mutated protein, as referred to herein, is a polypeptide homologous to a native RANKL protein, but which has an amino acid sequence different from the native protein because of one or a plurality of deletions, insertions or substitutions. The effect of any mutation made in a DNA encoding a mutated peptide may be easily determined by analyzing the ability of the mutated peptide to bind its counterstructure in a specific manner. Moreover, activity of RANKL analogs, muteins or

derivatives can be determined by any of the assays described herein (for example, induction of NF-kB activation).

Analogs of the inventive proteins may be constructed by, for example, making various substitutions of residues or sequences or deleting terminal or internal residues or sequences not needed for biological activity. For example, cysteine residues can be deleted or replaced with other amino acids to prevent formation of incorrect intramolecular disulfide bridges upon renaturation. Other approaches to mutagenesis involve modification of adjacent dibasic amino acid residues to enhance expression in yeast systems in which KEX2 protease activity is present.

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When a deletion or insertion strategy is adopted, the potential effect of the deletion or insertion on biological activity should be considered. Subunits of the inventive proteins may be constructed by deleting terminal or internal residues or sequences. Soluble forms of RANKL can be readily prepared and tested for their ability to induce NF-kB activation. Polypeptides corresponding to the cytoplasmic regions, and fragments thereof (for example, a death domain) can be prepared by similar techniques. Additional guidance as to the types of mutations that can be made is provided by a comparison of the sequence of RANKL to proteins that have similar structures, as well as by performing structural analysis of the inventive RANKL proteins.

Generally, substitutions should be made conservatively; i.e., the most preferred substitute amino acids are those which do not affect the biological activity of RANKL (i.e., ability of the inventive proteins to bind antibodies to the corresponding native protein in substantially equivalent a manner, the ability to bind the counterstructure in substantially the same manner as the native protein, the ability to induce a RANKL signal, or ability to induce NF-xB activation). Examples of conservative substitutions include substitution of amino acids outside of the binding domain(s) (either ligand/receptor or antibody binding areas for the extracellular domain, or regions that interact with other, intracellular proteins for the cytoplasmic domain), and substitution of amino acids that do not alter the secondary and/or tertiary structure of the native protein. Additional examples include substitutions of one polar residue for another, such as between Lys and Arg; Glu and Asp; or Gln and Asn. Other such conservative substitutions, for example, substitutions of entire regions having similar hydrophobicity characteristics, are well known.

Mutations in nucleotide sequences constructed for expression of analog proteins or fragments thereof must, of course, preserve the reading frame phase of the coding sequences and preferably will not create complementary regions that could hybridize to produce secondary mRNA structures such as loops or hairpins which would adversely affect translation of the mRNA.

Not all mutations in the nucleotide sequence which encodes a RANKL protein or fragments thereof will be expressed in the final product, for example, nucleotide substitutions may be made to enhance expression, primarily to avoid secondary structure loops in the transcribed mRNA (see EPA 75,444A, incorporated herein by reference), or to provide codons that are more readily translated by the selected host, e.g., the well-known *E. coli* preference codons for *E. coli* expression.

Although a mutation site may be predetermined, it is not necessary that the nature of the mutation *per se* be predetermined. For example, in order to select for optimum characteristics of mutants, random mutagenesis may be conducted and the expressed mutated proteins screened for the desired activity. Mutations can be introduced at particular loci by synthesizing oligonucleotides containing a mutant sequence, flanked by restriction sites enabling ligation to fragments of the native sequence. Following ligation, the resulting reconstructed sequence encodes an analog having the desired amino acid insertion, substitution, or deletion.

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Alternatively, oligonucleotide-directed site-specific mutagenesis procedures can be employed to provide an altered gene having particular codons altered according to the substitution, deletion, or insertion required. Exemplary methods of making the alterations set forth above are disclosed by Walder et al. (Gene 42:133, 1986); Bauer et al. (Gene 37:73, 1985); Craik (BioTechniques, January 1985, 12-19); Smith et al. (Genetic Engineering: Principles and Methods, Plenum Press, 1981); and U.S. Patent NOs. 4,518,584 and 4,737,462 disclose suitable techniques, and are incorporated by reference herein.

Additional embodiments of the inventive proteins include RANKL polypeptides encoded by DNAs capable of hybridizing to the DNAS of SEQ ID NO:10 or 12 under moderately stringent conditions (prewashing solution of 5 X SSC, 0.5% SDS, 1.0 mM EDTA (pH 8.0) and hybridization conditions of 50°C, 5 X SSC, overnight) to the DNA sequences encoding RANKL, or more preferably under stringent conditions (for example, hybridization in 6 X SSC at 63°C overnight; washing in 3 X SSC at 55°C), and other sequences which are degenerate to those which encode the RANKL. In one embodiment, RANKL polypeptides are at least about 70% identical in amino acid sequence to the amino acid sequence of native RANKL protein as set forth in SEQ ID NOs:10 and 12. In a preferred embodiment, RANKL polypeptides are at least about 80% identical in amino acid sequence to the native form of RANKL; most preferred polypeptides are those that are at least about 90% identical to native RANKL.

Percent identity may be determined using a computer program, for example, the GAP computer program described by Devereux et al. (*Nucl. Acids Res.* 12:387, 1984) and available from the University of Wisconsin Genetics Computer Group (UWGCG). For

fragments derived from the RANKL protein, the identity is calculated based on that portion of the RANKL protein that is present in the fragment

The biological activity of RANKL analogs or muteins can be determined by testing the ability of the analogs or muteins to induce a signal through RANK, for example, activation of transcription as described in the Examples herein. Alternatively, suitable assays, for example, an enzyme immunoassay or a dot blot, employing an antibody that binds native RANKL, or a soluble form of RANK, can be used to assess the activity of RANKL analogs or muteins. Suitable assays also include, for example, assays that measure the ability of a RANKL peptide or mutein to bind cells expressing RANK, and/or the biological effects thereon. Such methods are well known in the art.

Fragments of the RANKL nucleotide sequences are also useful. In one embodiment, such fragments comprise at least about 17 consecutive nucleotides, preferably at least about 25 nucleotides, more preferably at least 30 consecutive nucleotides, of the RANKL DNA disclosed herein. DNA and RNA complements of such fragments are provided herein, along with both single-stranded and double-stranded forms of the RANKL DNAs of SEQ ID NOs:10 and 12, and those encoding the aforementioned polypeptides. A fragment of RANKL DNA generally comprises at least about 17 nucleotides, preferably from about 17 to about 30 nucleotides. Such nucleic acid fragments (for example, a probe corresponding to the extracellular domain of RANKL) are used as a probe or as primers in a polymerase chain reaction (PCR).

The probes also find use in detecting the presence of RANKL nucleic acids in in vitro assays and in such procedures as Northern and Southern blots. Cell types expressing RANKL can be identified as well. Such procedures are well known, and the skilled artisan can choose a probe of suitable length, depending on the particular intended application. For PCR, 5' and 3' primers corresponding to the termini of a desired RANKL DNA sequence are employed to amplify that sequence, using conventional techniques.

Other useful fragments of the RANKL nucleic acids are antisense or sense oligonucleotides comprising a single-stranded nucleic acid sequence (either RNA or DNA) capable of binding to target RANKL mRNA (sense) or RANKL DNA (antisense) sequences. The ability to create an antisense or a sense oligonucleotide, based upon a cDNA sequence for a given protein is described in, for example, Stein and Cohen, Cancer Res. 48:2659, 1988 and van der Krol et al., BioTechniques 6:958, 1988.

## Uses of DNAs, Proteins and Analogs

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The RANKL DNAs, proteins and analogs described herein will have numerous uses, including the preparation of pharmaceutical compositions. For example, soluble forms of RANKL will be useful to transduce signal via RANK. RANKL compositions (both protein and DNAs) will also be useful in development of antibodies to RANKL, both

those that inhibit binding to RANK and those that do not. The inventive DNAs are useful for the expression of recombinant proteins, and as probes for analysis (either quantitative or qualitative) of the presence or distribution of RANKL transcripts.

The inventive proteins will also be useful in preparing kits that are used to detect soluble RANK or RANKL, or monitor RANK-related activity, for example, in patient specimens. RANKL proteins will also find uses in monitoring RANK-related activity in other samples or compositions, as is necessary when screening for antagonists or mimetics of this activity (for example, peptides or small molecules that inhibit or mimic, respectively, the interaction). A variety of assay formats are useful in such kits, including (but not limited to) ELISA, dot blot, solid phase binding assays (such as those using a biosensor), rapid format assays and bioassays.

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The purified RANKL according to the invention will facilitate the discovery of inhibitors of RANK, and thus, inhibitors of an inflammatory response (via inhibition of NF-kB activation). The use of a purified RANKL polypeptide in the screening for potential inhibitors is important and can virtually eliminate the possibility of interfering reactions with contaminants. Such a screening assay can utilize either the extracellular domain of RANKL, or a fragment thereof. Detecting the inhibiting activity of a molecule would typically involve use of a soluble form of RANKL derived from the extracellular domain in a screening assay to detect molecules capable of binding RANK and inhibiting binding of the RANKL.

In addition, RANKL polypeptides can also be used for structure-based design of RANKL-inhibitors. Such structure-based design is also known as "rational drug design." The RANKL polypeptides can be three-dimensionally analyzed by, for example, X-ray crystallography, nuclear magnetic resonance or homology modeling, all of which are well-known methods. The use of RANKL structural information in molecular modeling software systems to assist in inhibitor design is also encompassed by the invention. Such computer-assisted modeling and drug design may utilize information such as chemical conformational analysis, electrostatic potential of the molecules, protein folding, etc. A particular method of the invention comprises analyzing the three dimensional structure of RANKL for likely binding sites of substrates, synthesizing a new molecule that incorporates a predictive reactive site, and assaying the new molecule as described above.

Moreover, as shown in the Examples herein, soluble forms of RANKL will be useful to induce maturation of dendritic cells (DC), and to enhance their allo-stimulatory capacity. Accordingly, RANKL proteins will be useful in augmenting an immune response, and can be used for these purposes either ex vivo (i.e., in obtaining cells such as DC from an individual, exposing them to antigen and cytokines ex vivo, and readministering them to the individual) or in vivo (i.e., as a vaccine adjuvant that will augment humoral and/or cellular immunity). RANKL will also be useful promoting

viability of T cells in the presence of TGFB, which will also be helpful in regulating an immune response.

# Expression of Recombinant RANKL

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The proteins of the present invention are preferably produced by recombinant DNA methods by inserting a DNA sequence encoding RANKL protein or an analog thereof into a recombinant expression vector and expressing the DNA sequence in a recombinant expression system under conditions promoting expression. DNA sequences encoding the proteins provided by this invention can be assembled from cDNA fragments and short oligonucleotide linkers, or from a series of oligonucleotides, to provide a synthetic gene which is capable of being inserted in a recombinant expression vector and expressed in a recombinant transcriptional unit.

Recombinant expression vectors include synthetic or cDNA-derived DNA fragments encoding RANKL, or homologs, muteins or bioequivalent analogs thereof, operably linked to suitable transcriptional or translational regulatory elements derived from mammalian, microbial, viral or insect genes. Such regulatory elements include a transcriptional promoter, an optional operator sequence to control transcription, a sequence encoding suitable mRNA ribosomal binding sites, and sequences which control the termination of transcription and translation, as described in detail below. The ability to replicate in a host, usually conferred by an origin of replication, and a selection gene to facilitate recognition of transformants may additionally be incorporated.

DNA regions are operably linked when they are functionally related to each other. For example, DNA for a signal peptide (secretory leader) is operably linked to DNA for a polypeptide if it is expressed as a precursor which participates in the secretion of the polypeptide; a promoter is operably linked to a coding sequence if it controls the transcription of the sequence; or a ribosome binding site is operably linked to a coding sequence if it is positioned so as to permit translation. Generally, operably linked means contiguous and, in the case of secretory leaders, contiguous and in reading frame. DNA sequences encoding RANKL, or homologs or analogs thereof which are to be expressed in a microorganism will preferably contain no introns that could prematurely terminate transcription of DNA into mRNA.

Useful expression vectors for bacterial use can comprise a selectable marker and bacterial origin of replication derived from commercially available plasmids comprising genetic elements of the well known cloning vector pBR322 (ATCC 37017). Such commercial vectors include, for example, pKK223-3 (Pharmacia Fine Chemicals, Uppsala, Sweden) and pGEM1 (Promega Biotec, Madison, WI, USA). These pBR322 "backbone" sections are combined with an appropriate promoter and the structural sequence to be expressed. *E. coli* is typically transformed using derivatives of pBR322, a plasmid derived

from an E. coli species (Bolivar et al., Gene 2:95, 1977). pBR322 contains genes for ampicillin and tetracycline resistance and thus provides simple means for identifying transformed cells.

Promoters commonly used in recombinant microbial expression vectors include the β-lactamase (penicillinase) and lactose promoter system (Chang et al., Nature 275:615, 1978; and Goeddel et al., Nature 281:544, 1979), the tryptophan (trp) promoter system (Goeddel et al., Nucl. Acids Res. 8:4057, 1980; and EPA 36,776) and tac promoter (Maniatis, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, p. 412, 1982). A particularly useful bacterial expression system employs the phage λ PL promoter and cI857ts thermolabile repressor. Plasmid vectors available from the American Type Culture Collection which incorporate derivatives of the λ PL promoter include plasmid pHUB2, resident in E. coli strain JMB9 (ATCC 37092) and pPLc28, resident in E. coli RR1 (ATCC 53082).

Suitable promoter sequences in yeast vectors include the promoters for metallothionein, 3-phosphoglycerate kinase (Hitzeman et al., *J. Biol. Chem. 255*:2073, 1980) or other glycolytic enzymes (Hess et al., *J. Adv. Enzyme Reg. 7*:149, 1968; and Holland et al., *Biochem. 17*:4900, 1978), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase. Suitable vectors and promoters for use in yeast expression are further described in R. Hitzeman et al., EPA 73,657.

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Preferred yeast vectors can be assembled using DNA sequences from pBR322 for selection and replication in E. coli (Amp<sup>r</sup> gene and origin of replication) and yeast DNA sequences including a glucose-repressible ADH2 promoter and α-factor secretion leader. The ADH2 promoter has been described by Russell et al. (J. Biol. Chem. 258:2674, 1982) and Beier et al. (Nature 300:724, 1982). The yeast α-factor leader, which directs secretion of heterologous proteins, can be inserted between the promoter and the structural gene to be expressed. See, e.g., Kurjan et al., Cell 30:933, 1982; and Bitter et al., Proc. Natl. Acad. Sci. USA 81:5330, 1984. The leader sequence may be modified to contain, near its 3' end, one or more useful restriction sites to facilitate fusion of the leader sequence to foreign genes.

The transcriptional and translational control sequences in expression vectors to be used in transforming vertebrate cells may be provided by viral sources. For example, commonly used promoters and enhancers are derived from Polyoma, Adenovirus 2, Simian Virus 40 (SV40), and human cytomegalovirus. DNA sequences derived from the SV40 viral genome, for example, SV40 origin, early and late promoter, enhancer, splice, and polyadenylation sites may be used to provide the other genetic elements required for expression of a heterologous DNA sequence. The early and late promoters are particularly

useful because both are obtained easily from the virus as a fragment which also contains the SV40 viral origin of replication (Fiers et al., Nature 273:113, 1978). Smaller or larger SV40 fragments may also be used, provided the approximately 250 bp sequence extending from the Hind III site toward the BgII site located in the viral origin of replication is included. Further, viral genomic promoter, control and/or signal sequences may be utilized, provided such control sequences are compatible with the host cell chosen. Exemplary vectors can be constructed as disclosed by Oknyama and Berg (Mol. Cell. Biol. 3:280, 1983).

A useful system for stable high level expression of mammalian receptor cDNAs in C127 murine mammary epithelial cells can be constructed substantially as described by Cosman et al. (Mol. Immunol. 23:935, 1986). A preferred eukaryotic vector for expression of RANKL DNA is referred to as pDC406 (McMahan et al., EMBO J. 10:2821, 1991), and includes regulatory sequences derived from SV40, human iromunodeficiency virus (HIV), and Epstein-Barr virus (EBV). Other preferred vectors include pDC409 and pDC410, which are derived from pDC406. pDC410 was derived from pDC406 by substituting the EBV origin of replication with sequences encoding the SV40 large T antigen. pDC409 differs from pDC406 in that a Bgl II restriction site outside of the multiple cloning site has been deleted, making the Bgl II site within the multiple cloning site unique.

A useful cell line that allows for episomal replication of expression vectors, such as pDC406 and pDC409, which contain the EBV origin of replication, is CV-1/EBNA (ATCC CRL 10478). The CV-1/EBNA cell line was derived by transfection of the CV-1 cell line with a gene encoding Epstein-Barr virus nuclear antigen-1 (EBNA-1) and constitutively express EBNA-1 driven from human CMV immediate-early enhancer/promoter.

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#### Host Cells

Transformed host cells are cells which have been transformed or transfected with expression vectors constructed using recombinant DNA techniques and which contain sequences encoding the proteins of the present invention. Transformed host cells may express the desired protein (RANKL, or homologs or analogs thereof), but host cells transformed for purposes of cloning or amplifying the inventive DNA do not need to express the protein. Expressed proteins will preferably be secreted into the culture supernatant, depending on the DNA selected, but may be deposited in the cell membrane.

Suitable host cells for expression of proteins include prokaryotes, yeast or higher eukaryotic cells under the control of appropriate promoters. Prokaryotes include gram negative or gram positive organisms, for example *E. coli* or *Bacillus* spp. Higher eukaryotic cells include established cell lines of mammalian origin as described below. Cell-free translation systems could also be employed to produce proteins using RNAs

derived from the DNA constructs disclosed herein. Appropriate cloning and expression vectors for use with bacterial, fungal, yeast, and mammalian cellular hosts are described by Pouwels et al. (Cloning Vectors: A Laboratory Manual, Elsevier, New York, 1985), the relevant disclosure of which is hereby incorporated by reference.

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Prokaryotic expression hosts may be used for expression of RANKL, or homologs or analogs thereof that do not require extensive proteolytic and disulfide processing. Prokaryotic expression vectors generally comprise one or more phenotypic selectable markers, for example a gene encoding proteins conferring antibiotic resistance or supplying an autotrophic requirement, and an origin of replication recognized by the host to ensure amplification within the host. Suitable prokaryotic hosts for transformation include *E. coli, Bacillus subtilis, Salmonella typhimurium*, and various species within the genera *Pseudomonas, Streptomyces*, and *Staphylococcus*, although others may also be employed as a matter of choice.

Recombinant RANKL may also be expressed in yeast hosts, preferably from the Saccharomyces species, such as S. cerevisiae. Yeast of other genera, such as Pichia or Kluyveromyces may also be employed. Yeast vectors will generally contain an origin of replication from the 2µ yeast plasmid or an autonomously replicating sequence (ARS), promoter, DNA encoding the protein, sequences for polyadenylation and transcription termination and a selection gene. Preferably, yeast vectors will include an origin of replication and selectable marker permitting transformation of both yeast and E. coli, e.g., the ampicillin resistance gene of E. coli and S. cerevisiae trp1 gene, which provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan, and a promoter derived from a highly expressed yeast gene to induce transcription of a structural sequence downstream. The presence of the trp1 lesion in the yeast host cell genome then provides an effective environment for detecting transformation by growth in the absence of tryptophan.

Suitable yeast transformation protocols are known to those of skill in the art; an exemplary technique is described by Hinnen et al., *Proc. Natl. Acad. Sci. USA 75*:1929, 1978, selecting for Trp+ transformants in a selective medium consisting of 0.67% yeast nitrogen base, 0.5% casamino acids, 2% glucose, 10 µg/ml adenine and 20 µg/ml uracil. Host strains transformed by vectors comprising the ADH2 promoter may be grown for expression in a rich medium consisting of 1% yeast extract, 2% peptone, and 1% glucose supplemented with 80 µg/ml adenine and 80 µg/ml uracil. Derepression of the ADH2 promoter occurs upon exhaustion of medium glucose. Crude yeast supernatants are harvested by filtration and held at 4°C prior to further purification.

Various mammalian or insect cell culture systems can be employed to express recombinant protein. Baculovirus systems for production of heterologous proteins in insect cells are reviewed by Luckow and Summers, *Bio/Technology* 6:47 (1988). Examples of

suitable mammalian host cell lines include the COS-7 lines of monkey kidney cells, described by Gluzman (Cell 23:175, 1981), and other cell lines capable of expressing an appropriate vector including, for example, CV-1/EBNA (ATCC CRL 10478), L cells, C127, 3T3, Chinese hamster ovary (CHO), HeLa and BHK cell lines. Mammalian expression vectors may comprise nontranscribed elements such as an origin of replication, a suitable promoter and enhancer linked to the gene to be expressed, and other 5' or 3' flanking nontranscribed sequences, and 5' or 3' nontranslated sequences, such as necessary ribosome binding sites, a polyadenylation site, splice donor and acceptor sites, and transcriptional termination sequences.

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# Purification of Recombinant RANKL

Purified RANKL, and homologs or analogs thereof are prepared by culturing suitable host/vector systems to express the recombinant translation products of the DNAs of the present invention, which are then purified from culture media or cell extracts. For example, supernatants from systems which secrete recombinant protein into culture media can be first concentrated using a commercially available protein concentration filter, for example, an Amicon or Millipore Pellicon ultrafiltration unit.

Following the concentration step, the concentrate can be applied to a suitable purification matrix. For example, a suitable affinity matrix can comprise a counter structure protein or lectin or antibody molecule bound to a suitable support. Alternatively, an anion exchange resin can be employed, for example, a matrix or substrate having pendant diethylaminoethyl (DEAE) groups. The matrices can be acrylamide, agarose, dextran, cellulose or other types commonly employed in protein purification. Alternatively, a cation exchange step can be employed. Suitable cation exchangers include various insoluble matrices comprising sulfopropyl or carboxymethyl groups. Sulfopropyl groups are preferred. Gel filtration chromatography also provides a means of purifying the inventive proteins.

Affinity chromatography is a particularly preferred method of purifying RANKL and homologs thereof. For example, a RANKL expressed as a fusion protein comprising an immunoglobulin Fc region can be purified using Protein A or Protein G affinity chromatography. Moreover, a RANKL protein comprising an oligomerizing zipper domain may be purified on a resin comprising an antibody specific to the oligomerizing zipper domain. Monoclonal antibodies against the RANKL protein may also be useful in affinity chromatography purification, by utilizing methods that are well-known in the art. A ligand may also be used to prepare an affinity matrix for affinity purification of RANKL.

Finally, one or more reversed-phase high performance liquid chromatography (RP-HPLC) steps employing hydrophobic RP-HPLC media, e.g., silica gel having pendant methyl or other aliphatic groups, can be employed to further purify a RANKL composition.

Some or all of the foregoing purification steps, in various combinations, can also be employed to provide a homogeneous recombinant protein.

Recombinant protein produced in bacterial culture is usually isolated by initial extraction from cell pellets, followed by one or more concentration, salting-out, aqueous ion exchange or size exclusion chromatography steps. Finally, high performance liquid chromatography (HPLC) can be employed for final purification steps. Microbial cells employed in expression of recombinant protein can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents.

Fermentation of yeast which express the inventive protein as a secreted protein greatly simplifies purification. Secreted recombinant protein resulting from a large-scale fermentation can be purified by methods analogous to those disclosed by Urdal et al. (*J. Chromatog.* 296:171, 1984). This reference describes two sequential, reversed-phase HPLC steps for purification of recombinant human GM-CSF on a preparative HPLC column.

Protein synthesized in recombinant culture is characterized by the presence of cell components, including proteins, in amounts and of a character which depend upon the purification steps taken to recover the inventive protein from the culture. These components ordinarily will be of yeast, prokaryotic or non-human higher eukaryotic origin and preferably are present in innocuous contaminant quantities, on the order of less than about 1 percent by weight. Further, recombinant cell culture enables the production of the inventive proteins free of other proteins which may be normally associated with the proteins as they are found in nature in the species of origin.

# 25 Uses and Administration of RANKL Compositions

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The present invention provides methods of using therapeutic compositions comprising an effective amount of a protein and a suitable diluent and carrier, and methods for regulating an immune or inflammatory response. The use of RANKL in conjunction with soluble cytokine receptors or cytokines, or other immunoregulatory molecules is also contemplated.

For therapeutic use, purified protein is administered to a patient, preferably a human, for treatment in a manner appropriate to the indication. Thus, for example, RANKL protein compositions administered to regulate immune function can be given by bolus injection, continuous infusion, sustained release from implants, or other suitable technique. Typically, a therapeutic agent will be administered in the form of a composition comprising purified RANKL, in conjunction with physiologically acceptable carriers, excipients or diluents. Such carriers will be nontoxic to recipients at the dosages and concentrations employed.

Ordinarily, the preparation of such protein compositions entails combining the inventive protein with buffers, antioxidants such as ascorbic acid, low molecular weight (less than about 10 residues) polypeptides, proteins, amino acids, carbohydrates including glucose, sucrose or dextrins, chelating agents such as EDTA, glutathione and other stabilizers and excipients. Neutral buffered saline or saline mixed with conspecific serum albumin are exemplary appropriate diluents. Preferably, product is formulated as a lyophilizate using appropriate excipient solutions (e.g., sucrose) as diluents. Appropriate dosages can be determined in trials. The amount and frequency of administration will depend, of course, on such factors as the nature and severity of the indication being treated, the desired response, the condition of the patient, and so forth.

As shown hrein, RANKL has beneficial effects on various cells important in the immune system. Accordingly, RANKL may be adminstered to an individual as a vaccine adjuvant, or as a therapeutic agent to upregulate an immune resposne, for example, ininfectious disease. Moreover, NF-KB has been found to play a protective role in preventing apoptotic death of cells induced by TNF-α or chemotherapy. Accordingly, agonists of RANK (i.e., RANKL and agonistic antibodies) will be useful in protecting . . . . RANK-expressing cells from the negative effects of chemotherapy or the presence of high levels of TNF-α such as occur in sepsis (see, i.e., Barinaga, Science 274"724, 1996, and the articles by Beg and Baltimore and Wang etal., pages 782 and 784 of that same issue of

Throughout the description and claims of this specification, the word . "comprise" and variations of the word, such as "comprising" and comprises", is not intended to exclude other additives, components, integers or steps".

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The following examples are offered by way of illustration, and not by way of limitation. Those skilled in the art will recognize that variations of the invention embodied in the examples can be made, especially in light of the teachings of the ..... various references cited herein, the disclosures of which are incorporated by reference.

# EXAMPLE 1

30 The example describes the identification and isolation of a DNA encoding a novel member of the TNF receptor superfamily. A partial cDNA insert with a predicted open reading frame having some similarity to CD40 (a cell-surface antigen present on the surface of both normal and neoplastic human B cells that has been shown to play an important role in B-cell proliferation and differentiation; Starnenkovic et al., EMBO J. 8:1403, 1989), was identified in a database containing sequence information from cDNAs generated from human bone marrow-derived dendritic cells (DC). The insert was excised from the vector by restriction endonuclease digestion, gel purified. labeled with 32P, and used to hybridize to colony blots generated from a DC cDNA library containing larger cDNA inserts using

high stringency hybridization and washing techniques (hybridization in 5xSSC, 50% formamide at 42°C overnight, washing in 0.5xSSC at 63°C); other suitable high stringency conditions are disclosed in Sambrook et al. in Molecular Cloning: A Laboratory Manual, 2nd ed. (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY; 1989), 9.52-9.55. Initial experiments yielded a clone referred to as 9D-8A (SEQ ID NO:1); subsequent analysis indicated that this clone contained all but the extreme 5' end of a novel cDNA, with predicted intron sequence at the extreme 5' end (nucleotides 1-92 of SEQ ID NO:1). Additional colony hybridizations were performed, and a second clone was isolated. The second clone, referred to as 9D-15C (SEQ ID NO:3), contained the 5' end without intron interruption but not the full 3'end. SEQ ID NO:5 shows the nucleotide and amino acid sequence of a predicted full-length protein based on alignment of the overlapping sequences of SEQ ID NO:1 and 3.

The encoded protein was designated RANK, for receptor activator of NF-kB. The cDNA encodes a predicted Type 1 transmembrane protein having 616 amino acid residues, with a predicted 24 amino acid signal sequence (the computer predicted cleavage site is after Leu24), a 188 amino acid extracellular domain, a 21 amino acid transmembrane domain, and a 383 amino acid cytoplasmic tail. The extracellular region of RANK displayed significant amino acid homology (38.5% identity, 52.3% similarity) to CD40. A cloning vector (pBluescriptSK-) containing human RANK sequence, designated pBluescript:huRANK (in E coli DH10B), was deposited with the American Type Culture Collection, Rockville, MD (ATCC) on December 20, 1996, under terms of the Budapest Treaty, and given accession number 98285.

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## **EXAMPLE 2**

This example describes construction of a RANK DNA construct to express a RANK/Fc fusion protein. A soluble form of RANK fused to the Fc region of human IgG<sub>1</sub> was constructed in the mammalian expression vector pDC409 (USSN 08/571,579). This expression vector encodes the leader sequence of the Cytomegalovirus (CMV) open reading frame R27080 (SEQ ID NO:9), followed by amino acids 33-213 of RANK, followed by a mutated form of the constant domain of human IgG<sub>1</sub> that exhibits reduced affinity for Fc receptors (SEQ ID NO:8; for the fusion protein, the Fc portion of the construct consisted of Arg3 through Lys232). An alternative expression vector encompassing amino acids 1-213 of RANK (using the native leader sequence) followed by the IgG<sub>1</sub> mutein was also prepared. Both expression vectors were found to induce high levels of expression of the RANK/Fc fusion protein in transfected cells.

To obtain RANK/Fc protein, a RANK/Fc expression plasmid is transfected into CV-1/EBNA cells, and supernatants are collected for about one week. The RANK/Fc fusion protein is purified by means well-known in the art for purification of Fc fusion

proteins, for example, by protein A sepharose column chromatography according to manufacturer's recommendations (i.e., Pharmacia, Uppsala, Sweden). SDS-polyacrylamide gel electrophoresis analysis indicted that the purified RANK/Fc protein migrated with a molecular weight of ~55kDa in the presence of a reducing agent, and at a molecular weight of ~110kDa in the absence of a reducing agent.

N-terminal amino acid sequencing of the purified protein made using the CMV R27080 leader showed 60% cleavage after Ala20, 20% cleavage after Pro22 and 20% cleavage after Arg28 (which is the Furin cleavage site; amino acid residues are relative to SEQ ID NO:9); N-terminal amino acid analysis of the fusion protein expressed with the native leader showed cleavage predominantly after Gln25 (80% after Gln25 and 20% after Arg23; amino acid residues are relative to SEQ ID NO:6, full-length RANK). Both fusion proteins were able to bind a ligand for RANK is a specific manner (i.e., they bound to the surface of various cell lines such as a murine thymoma cell line, ELA), indicating that the presence of additional amino acids at the N-terminus of RANK does not interfere with its ability to bind RANKL. Moreover, the construct comprising the CMV leader encoded RANK beginning at amino acid 33; thus, a RANK peptide having an N-terminus at an amino acid between Arg23 and Pro33, inclusive, is expected to be able to bind a ligand for RANK in a specific manner.

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Other members of the TNF receptor superfamily have a region of amino acids between the transmembrane domain and the ligand binding domain that is referred to as a 'spacer' region, which is not necessary for ligand binding. In RANK, the amino acids between 196 and 213 are predicted to form such a spacer region. Accordingly, a soluble form of RANK that terminates with an amino acid in this region is expected to retain the ability to bind a ligand for RANK in a specific manner. Preferred C-terminal amino acids for soluble RANK peptides are selected from the group consisting of amino acids 213 and 196 of SEQ ID NO:6, although other amino acids in the spacer region may be utilized as a C-terminus.

## EXAMPLE 3

This example illustrates the preparation of monoclonal antibodies against RANK. Preparations of purified recombinant RANK, for example, or transfected cells expressing high levels of RANK, are employed to generate monoclonal antibodies against RANK using conventional techniques, such as those disclosed in U.S. Patent 4,411,993. DNA encoding RANK can also be used as an immunogen, for example, as reviewed by Pardoll and Beckerleg in *Immunity* 3:165, 1995. Such antibodies are likely to be useful in interfering with RANK-induced signaling (antagonistic or blocking antibodies) or in inducing a signal by cross-linking RANK (agonistic antibodies), as components of

#### EXAMPLE 4

This example illustrates the induction of NF-kB activity by RANK in 293/EBNA cells (cell line was derived by transfection of the 293 cell line with a gene encoding Epstein-Barr virus nuclear antigen-1 (EBNA-1) that constitutively express EBNA-1 driven from human CMV immediate-early enhancer/promoter). Activation of NF-kB activity was measured in 293/EBNA cells essentially as described by Yao et al. (Immunity 3:811, 1995). Nuclear extracts were prepared and analyzed for NF-kB activity by a gel retardation assay using a 25 base pair oligonucleotide spanning the NF-kB binding sites. Two million cells were seeded into 10 cm dishes two days prior to DNA transfection and cultured in DMEM-F12 media containing 2.5% FBS (fetal bovine serum). DNA transfections were performed as described herein for the IL-8 promoter/reporter assays.

Nuclear extracts were prepared by solubilization of isolated nuclei with 400 mM NaCl (Yao et al., supra). Oligonucleotides containing an NF-kB binding site were annealed and endlabeled with <sup>32</sup>P using T4 DNA polynucleotide kinase. Mobility shift reactions contained 10 µg of nuclear extract, 4 µg of poly(dI-dC) and 15,000 cpm labeled double-stranded oligonucleotide and incubated at room temperature for 20 minutes. Resulting protein-DNA complexes were resolved on a 6% native polyacrylamide gel in 0.25 X Tris-borate-EDTA buffer.

Overexpression of RANK resulted in induction of NF-kB activity as shown by an appropriate shift in the mobility of the radioactive probe on the gel. Similar results were observed when RANK was triggered by a ligand that binds RANK and transduces a signal to cells expressing the receptor (i.e., by co-transfecting cells with human RANK and murine RANKL DNA; see Example 7 below), and would be expected to occur when triggering is done with agonistic antibodies.

EXAMPLE 5

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This example describes a gene promoter/reporter system based on the human Interleukin-8 (IL-8) promoter used to analyze the activation of gene transcription in vivo. The induction of human IL-8 gene transcription by the cytokines Interleukin-1 (IL-1) or tumor necrosis factor-alpha (TNF-α) is known to be dependent upon intact NF-κB and NF-IL-6 transcription factor binding sites. Fusion of the cytokine-responsive IL-8 promoter with a cDNA encoding the murine IL-4 receptor (mIL-4R) allows measurement of promoter activation by detection of the heterologous reporter protein (mIL-4R) on the cell surface of transfected cells.

Human kidney epithelial cells (293/EBNA) are transfected (via the DEAE/DEXTRAN method) with plasmids encoding: 1). the reporter/promoter construct (referred to as pIL-8rep), and 2), the cDNA(s) of interest. DNA concentrations are always kept constant by the addition of empty vector DNA. The 293/EBNA cells are plated at a

density of 2.5 x 10<sup>4</sup> cells/ml (3 ml/ well) in a 6 well plate and incubated for two days prior to transfection. Two days after transfection, the mIL-4 receptor is detected by a radioimmunoassay (RIA) described below.

In one such experiment, the 293/EBNA cells were co-transfected with DNA encoding RANK and with DNA encoding RANKL (see Example 7 below). Co-expression of this receptor and its counterstructure by cells results in activation of the signaling process of RANK. For such co-transfection studies, the DNA concentration/well for the DEAE transfection were as follows: 40 ng of pIL-8rep [pBluescriptSK- vector (Stratagene)]; 0.4 ng CD40 (DNA encoding CD40, a control receptor; pCDM8 vector); 0.4 ng RANK (DNA encoding RANK; pDC409 vector), and either 1-50 ng CD40L (DNA encoding the ligand for CD40, which acts as a positive control when co-transfected with CD40 and as a negative control when co-transfected with RANK; in pDC304) or RANKL (DNA encoding a ligand for RANK; in pDC406). Similar experiments can be done using soluble RANKL or agonistic antibodies to RANK to trigger cells transfected with RANK.

For the mIL-4R-specific RIA, a monoclonal antibody reactive with mIL-4R is labeled with <sup>125</sup>I via a Chloramine T conjugation method; the resulting specific activity is typically 1.5 x 10<sup>16</sup> cpm/nmol. After 48 hours, transfected cells are washed once with media (DMEM/F12 5% FBS). Non-specific binding sites are blocked by the addition of pre-warmed binding media containing 5% non-fat dry milk and incubation at 37°C/5% CO<sub>2</sub> in a tissue culture incubator for one hour. The blocking media is decanted and binding buffer containing <sup>125</sup>I anti-tnIL-4R (clone MI; rat IgGI) is added to the cells and incubated with rocking at room temperature for 1 hour. After incubation of the cells with the radio-labeled antibody, cells are washed extensively with binding buffer (2X) and twice with phosphate-buffered saline (PBS). Cells are lysed in 1 ml of 0.5M NaOH, and total radioactivity is measured with a gamma counter.

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Using this assay, 293/EBNA co-transfected with DNAs encoding RANK demonstrated transcriptional activation, as shown by detection of mull\_4R on the cell surface. Overexpression of RANK resulted in transcription of mull\_4R, as did triggering of the RANK by RANKL. Similar results are observed when RANK is triggered by agonistic antibodies.

## **EXAMPLE 6**

This example illustrates the association of RANK with TRAF proteins. Interaction of RANK with cytoplasmic TRAF proteins was demonstrated by co-immunoprecipitation assays essentially as described by Hsu et al. (*Cell* 84:299; 1996). Briefly, 293/EBNA cells were co-transfected with plasmids that direct the synthesis of RANK and epitope-tagged (FLAG®; SEQ ID NO:7) TRAF2 or TRAF3. Two days after transfection, surface proteins

were labeled with biotin-ester, and cells were lysed in a buffer containing 0.5% NP-40. RANK and proteins associated with this receptor were immunoprecipitated with anti-RANK, washed extensively, resolved by electrophoretic separation on a 6-10% SDS polyacrylamide gel and electrophoretically transferred to a nitrocellulose membrane for Western blotting. The association of TRAF2 and TRAF3 proteins with RANK was visualized by probing the membrane with an antibody that specifically recognizes the FLAG® epitope. TRAFs 2 and 3 did not immunoprecipitate with anti-RANK in the absence of RANK expression.

## EXAMPLE 7

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This example describes isolation of a ligand for RANK, referred to as RANKL, by direct expression cloning. The ligand was cloned essentially as described in USSN 08/249,189, filed May 24, 1994 (the relevant disclosure of which is incorporated by reference herein), for CD40L. Briefly, a library was prepared from a clone of a mouse thymoma cell line EL-4 (ATCC TIB 39), called EL-40.5, derived by sorting five times with biotinylated CD40/Fc fusion protein in a FACS (fluorescence activated cell sorter). The cDNA library was made using standard methodology; the plasmid DNA was isolated and transfected into sub-confluent CV1-EBNA cells using a DEAE-dextran method. Transfectants were screened by slide autoradiography for expression of RANKL using a two-step binding method with RANK/Fc fusion protein as prepared in Example 2 followed by radioiodinated goat anti-human IgG antibody.

A clone encoding a protein that specifically bound RANK was isolated and sequenced; the clone was referred to as 11H. An expression vector containing murine RANKL sequence, designated pDC406:muRANK-L (in *E. coli* DH10B), was deposited with the American Type Culture Collection, Rockville, MD (ATCC) on December 20, 1996, under terms of the Budapest Treaty, and given accession number 98284. The nucleotide sequence and predicted amino acid sequence of this clone are illustrated in SEQ ID NO:10. This clone did not contain an initiator methionine; additional, full-length clones were obtained from a 7B9 library (prepared substantially as described in US patent 5,599,905, issued February 4, 1997); the 5' region was found to be identical to that of human RANKL as shown in SEQ ID NO: 12, amino acids 1 through 22, except for substitution of a Gly for a Thr at residue 9.

This ligand is useful for assessing the ability of RANK to bind RANKL by a number of different assays. For example, transfected cells expressing RANKL can be used in a FACS assay (or similar assay) to evaluate the ability of soluble RANK to bind RANKL. Moreover, soluble forms of RANKL can be prepared and used in assays that are known in the art (i.e., ELISA or BIAcore assays essentially as described in USSN 08/249,189, filed May 24, 1994). RANKL is also useful in affinity purification of RANK.

and as a reagent in methods to measure the levels of RANK in a sample. Soluble RANKL is also useful in inducing NF-kB activation and thus protecting cells that express RANK from apoptosis.

#### **EXAMPLE 8**

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This example describes the isolation of a human RANK ligand (RANKL) using a PCR-based technique. Murine RANK ligand-specific oligonucleotide primers were used in PCR reactions using human cell line-derived first strand cDNAs as templates. Primers corresponded to nucleotides 478-497 and to the complement of nucleotides 858-878 of murine RANK ligand (SEQ ID NO:10). An amplified band approximately 400 bp in length from one reaction using the human epidermoid cell line KB (ATCC CCL-17) was gel purified, and its nucleotide sequence determined; the sequence was 85% identical to the corresponding region of murine RANK ligand, confirming that the fragment was from human RANKL.

To obtain full-length human RANKL cDNAs, two human RANKL-specific oligonucleotides derived from the KB PCR product nucleotide sequence were radiolabeled and used as hybridization probes to screen a human PBL cDNA library prepared in lambda gt10 (Stratagene, La Jolla, CA), substantially as described in US patent 5,599,905, issued February 4, 1997. Several positive hybridizing plaques were identified and purified, their inserts subcloned into pBluescript SK<sup>-</sup> (Stratagene, La Jolla, CA), and their nucleotide sequence determined. One isolate, PBL3, was found to encode most of the predicted human RANKL, but appeared to be missing approximately 200 bp of 5' coding region. A second isolate, PBL5 was found to encode much of the predicted human RANKL, including the entire 5' end and an additional 200 bp of 5' untranslated sequence.

The 5' end of PBL5 and the 3' end of PBL3 were ligated together to form a full length cDNA encoding human RANKL. The nucleotide and predicted amino acid sequence of the full-length human RANK ligand is shown in SEQ ID NO:12. Human RANK ligand shares 83% nucleotide and 84% amino acid identity with murine RANK ligand. A plasmid vector containing human RANKL sequence, designated pBluescript:huRANK-L (in E. coli DH10B), was deposited with the American Type Culture Collection, Rockville, MD (ATCC) on March 11, 1997 under terms of the Budapest Treaty, and given accession number 98354.

Murine and human RANKL are Type 2 transmembrane proteins. Murine RANKL contains a predicted 48 amino acid intracellular domain, 21 amino acid transmembrane domain and 247 amino acid extracellular domain. Human RANKL contains a predicted 47 amino acid intracellular domain, 21 amino acid transmembrane domain and 249 amino acid extracellular domain.

# EXAMPLE 9

This example describes the chromosomal mapping of human RANK using PCR-based mapping strategies. Initial human chromosomal assignments were made using RANK and RANKL-specific PCR primers and a BIOS Somatic Cell Hybrid PCRable DNA kit from BIOS Laboratories (New Haven, CT), following the manufacturer's instructions. RANK mapped to human chromosome 18; RANK ligand mapped to human chromosome 13. More detailed mapping was performed using a radiation hybrid mapping panel Genebridge 4 Radiation Hybrid Panel (Research Genetics, Huntsville, AL; described in Walter, MA et al., Nature Genetics 7:22-28, 1994). Data from this analysis was then submitted electronically to the MIT Radiation Hybrid Mapper (URL: http://www-genome.wi.mit.edu/cgi-bin/contig/rhmapper.pl) following the instructions contained therein. This analysis yielded specific genetic marker names which, when submitted electronically to the NCBI Entrez browser (URL: http://www3.ncbi.nlm.nih.gov/htbin-post/Entrez/query?db=c&form=0), yielded the specific map locations. RANK mapped to chromosome 18q22.1, and RANKL mapped to chromosome 13q14.

#### **EXAMPLE 10**

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This example illustrates the preparation of monoclonal antibodies against RANKL. Preparations of purified recombinant RANKL, for example, or transfixed cells expressing high levels of RANKL, are employed to generate monoclonal antibodies against RANKL using conventional techniques, such as those disclosed in US Patent 4,411,993. DNA encoding RANKL can also be used as an immunogen, for example, as reviewed by Pardoll and Beckerleg in *Immunity* 3:165, 1995. Such antibodies are likely to be useful in interfering with RANKL signaling (antagonistic or blocking antibodies), as components of diagnostic or research assays for RANKL or RANKL activity, or in affinity purification of RANKL.

To immunize rodents, RANKL immunogen is emulsified in an adjuvant (such as complete or incomplete Freund's adjuvant, alum, or another adjuvant, such as Ribi adjuvant R700 (Ribi, Hamilton, MT), and injected in amounts ranging from 10-100 µg subcutaneously into a selected rodent, for example, BALB/c mice or Lewis rats. DNA may be given intradermally (Raz et al., *Proc. Natl. Acad. Sci. USA* 91:9519, 1994) or intamuscularly (Wang et al., *Proc. Natl. Acad. Sci. USA* 90:4156, 1993); saline has been found to be a suitable diluent for DNA-based antigens. Ten days to three weeks days later, the immunized animals are boosted with additional immunogen and periodically boosted thereafter on a weekly, biweekly or every third week immunization schedule.

Serum samples are periodically taken by retro-orbital bleeding or tail-tip excision for testing by dot-blot assay (antibody sandwich), ELISA (enzyme-linked immunosorbent

assay), immunoprecipitation, or other suitable assays, including FACS analysis. Following detection of an appropriate antibody titer, positive animals are given an intravenous injection of antigen in saline. Three to four days later, the animals are sacrificed, splenocytes harvested, and fused to a murine myeloma cell line (e.g., NS1 or preferably Ag 8.653 [ATCC CRL 1580]). Hybridoma cell lines generated by this procedure are plated in multiple microtiter plates in a selective medium (for example, one containing hypoxanthine, aminopterin, and thymidine, or HAT) to inhibit proliferation of non-fused cells, myeloma-myeloma hybrids, and splenocyte-splenocyte hybrids.

Hybridoma clones thus generated can be screened by ELISA for reactivity with RANKL, for example, by adaptations of the techniques disclosed by Engvall et al., *Immunochem.* 8:871 (1971) and in US Patent 4,703,004. A preferred screening technique is the antibody capture technique described by Beckman et al., *J. Immunol.* 144:4212 (1990). Positive clones are then injected into the peritoneal cavities of syngeneic rodents to produce ascites containing high concentrations (>1 mg/ml) of anti-RANK monoclonal antibody. The resulting monoclonal antibody can be purified by ammonium sulfate precipitation followed by gel exclusion chromatography. Alternatively, affinity chromatography based upon binding of antibody to protein A or protein G can also be used, as can affinity chromatography based upon binding to RANKL protein. Using the methods described herein to monitor the activity of the mAbs, both blocking (i.e., antibodies that bind RANKL and inhibit binding to RANK) and non-blocking (i.e., antibodies that bind RANKL and do not inhibit binding) are isolated.

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#### **EXAMPLE 11**

This example demonstrates that RANK expression can be up-regulated. Human peripheral blood T cells were purified by flow cytometry sorting or by negative selection using antibody coated beads, and activated with anti-CD3 (OKT3, Dako) coated plates or phytohemagglutinin in the presence or absence of various cytokines, including Interleukin-4 (IL-4), Transforming Growth Factor-β (TGF-β) and other commercially available cytokines (IL1-α, IL-2, IL-3, IL-6, IL-7, IL-8, IL-10, IL-12, IL-15, IFN-γ, TNF- α). Expression of RANK was evaluated by FACS in a time course experiment for day 2 to day 8, using a mouse monoclonal antibody mAb144 (prepared as described in Example 3), as shown in the table below. Results are expressed as '+' to '++++' referring to the relative increase in intensity of staining with anti-RANK. Double labeling experiments using both anti-RANK and anti-CD8 or anti-CD4 antibodies were also performed.

Table 1: Upregulation of RANK by Cytokines

Cytokine (concentration)	Results:
IL-4 (50 ng/ml)	+
TGF-ß (5 ng/ml)	+ to ++
IL-4 (50 ng/ml) +TGF-8 (5 ng/ml)	++++
IL1-α (10ng/ml)	-
IL-2 (20ng/ml)	-
IL-3 (25ng/ml)	-
IL-7 (20ng/ml)	-
IL-8 (10ng/ml)	-
IL-10 (50ng/ml)	_
IL-12 (10ng/ml)	-
lL-15 (10ng/ml)	-
IFN-γ (100U/m!)	-
TNF-α (10ng/ml)	•

Of the cytokines tested, IL-4 and TGF-8 increased the level of RANK expression on both CD8+ cytotoxic and CD4+ helper T cells from day 4 to day 8. The combination of IL-4 and TGF-8 acted synergistically to upregulate expression of this receptor on activated T cells. This particular combination of cytokines is secreted by suppresser T cells, and is believed to be important in the generation of tolerance (reviewed in Mitchison and Sieper, Z. Rheumatol. 54:141, 1995), implicating the interaction of RANK in regulation of an immune response towards either tolerance or induction of an active immune response.

### EXAMPLE 12

This example illustrates the influence of RANK.Fc and hRANKL on activated T cell growth. The addition of TGFB to anti-CD3 activated human peripheral blood T lymphocytes induces proliferation arrest and ultimately death of most lymphocytes within the first few days of culture. We tested the effect of RANK:RANKL interactions on TGFB-treated T cells by adding RANK.Fc or soluble human RANKL to T cell cultures.

Human peripheral blood T cells (7 x 10<sup>5</sup> PBT) were cultured for six days on anti-CD3 (OKT3, 5µg/ml) and anti-Flag (M1, 5µg/ml) coated 24 well plates in the presence of TGF8 (lng/ml) and IL-4 (10ng/ml), with or without recombinant FLAG-tagged soluble hRANKL (1µg/ml) or RANK.Fc (10µg/ml). Viable T cell recovery was determined by triplicate trypan blue countings.

The addition of RANK.Fc significantly reduced the number of viable T cells recovered after six days, whereas soluble RANKL greatly increased the recovery of viable T cells (Figure 1). Thus, endogenous or exogenous RANKL enhances the number of viable T cells generated in the presence of TGF8. TGF8, along with IL-4, has been implicated in immune response regulation when secreted by the T<sub>H</sub>3/regulatory T cell subset. These T cells are believed to mediate bystander suppression of effector T cells. Accordingly, RANK and its ligand may act in an auto/paracrine fashion to influence T cell tolerance. Moreover, TGF8 is known to play a role in the evasion of the immune system effected by certain pathogenic or opportunistic organisms. In addition to playing a role in the development of tolerance, RANK may also play a role in immune system evasion by pathogens.

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# EXAMPLE 13

This example illustrates the influence of the interaction of RANK on CD1a<sup>+</sup> dendritic cells (DC). Functionally mature dendritic cells (DC) were generated *in vitro* from CD34+ bone marrow (BM) progenitors. Briefly, human BM cells from normal healthy volunteers were density fractionated using Ficoll medium and CD34+ cells immunoaffinity isolated using an anti-CD34 matrix column (Ceprate, CellPro). The CD34+ BM cells were then cultured in human GM-CSF (20 ng/ml), human IL-4 (20 ng/ml), human TNF-α (20 ng/ml), human CHO-derived Fl3L (FL; 100 ng/ml) in Super McCoy's medium supplemented with 10% fetal calf serum in a fully humidified 37°C incubator (5% CO<sub>2</sub>) for 14 days. CD1a<sup>+</sup>, HLA-DR+ DC were then sorted using a FACStar Plus<sup>TM</sup>, and used for biological evaluation of RANK

On human CD1a+DC derived from CD34+ bone marrow cells, only a subset (20-30%) of CD1a+DC expressed RANK at the cell surface as assessed by flow cytometric

analysis. However, addition of CD40L to the DC cultures resulted in RANK surface expression on the majority of CD1a<sup>+</sup> DC. CD40L has been shown to activate DC by enhancing *in vitro* cluster formation, inducing DC morphological changes and upregulating HLA-DR, CD54, CD58, CD80 and CD86 expression

Addition of RANKL to DC cultures significantly increased the degree of DC aggregation and cluster formation above control cultures, similar to the effects seen with CD40L (Figure 2). Sorted human CD1a+ DC were cultured in a cytokine cocktail (GM-CSF, IL-4, TNF-α and FL) (upper left panel), in cocktail plus CD40L (lμg/ml) (upper right), in cocktail plus RANKL (lμg/ml) (lower left), or in cocktail plus heat inactivated (ΔΗ) RANKL (lμg/ml) (lower right) in 24-well flat bottomed culture plates in 1 ml culture media for 48-72 hours and then photographed using an inversion microscope. An increase in DC aggregation and cluster formation above control cultures was not evident when heat inactivated RANKL was used, indicating that this effect was dependent on biologically active protein. However, initial phenotypic analysis of adhesion molecule expression indicated that RANKL-induced clustering was not due to increased levels of CD2, CD11a, CD54 or CD58.

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The addition of RANKL to CD1a<sup>+</sup> DC enhanced their allo-stimulatory capacity in a mixed lymphocyte reaction (MLR) by at least 3- to 10-fold, comparable to CD40L-cultured DC (Figure 3). Allogeneic T cells (1x105) were incubated with varying numbers of irradiated (2000 rad) DC cultured as indicated above for Figure 2 in 96-well round bottomed culture plates in 0.2 ml culture medium for four days. The cultures were pulsed with 0.5 mCi [3H]-thymidine for eight hours and the cells harvested onto glass fiber sheets for counting on a gas phase  $\beta$  counter. The background counts for either T cells or DC cultured alone were <100 cpm. Values represent the mean  $\pm$  SD of triplicate cultures. Heat inactivated RANKL had no effect. DC allo-stimulatory activity was not further enhanced when RANKL and CD40L were used in combination, possibly due to DC functional capacity having reached a maximal level with either cytokine alone. Neither RANKL nor CD40L enhanced the *in vitro* growth of DC over the three day culture period. Unlike CD40L, RANKL did not significantly increase the levels of HLA-DR expression nor the expression of CD80 or CD86.

RANKL can enhance DC cluster formation and functional capacity without modulating known molecules involved in cell adhesion (CD18, CD54), antigen presentation (HLA-DR) or costimulation (CD86), all of which are regulated by CD40/CD40L signaling. The lack of an effect on the expression of these molecules suggests that RANKL may regulate DC function via an alternate pathway(s) distinct from CD40/CD40L. Given that CD40L regulates RANK surface expression on in vitrogenerated DC and that CD40L is upregulated on activated T cells during DC-T cell

interactions, RANK and its ligand may form an important part of the activation cascade that is induced during DC-mediated T cell expansion. Furthermore, culture of DC in RANKL results in decreased levels of CDlb/c expression, and increased levels of CD83. Both of these molecules are similarly modulated during DC maturation by CD40L (Caux et al. J. Exp. Med. 180:1263; 1994), indicating that RANKL induces DC maturation.

Dendritic cells are referred to as "professional" antigen presenting cells, and have a high capacity for sensitizing MHC-restricted T cells. There is growing interest in using dendritic cells ex vivo as tumor or infectious disease vaccine adjuvants (see, for example, Romani, et al., J. Exp. Med., 180:83, 1994). Therefore, an agent such as RANKL that induces DC maturation and enhances the ability of dendritic cells to stimulate an immune response is likely to be useful in immunotherapy of various diseases.

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#### **EXAMPLE 14**

This example describes the isolation of the murine homolog of RANK, referred to as muRANK. MuRANK was isolated by a combination of cross-species PCR and colony hybridization. The conservation of Cys residues in the Cys-rich pseudorepeats of the extracellular domains of TNFR superfamily member proteins was exploited to design human RANK-based PCR primers to be used on murine first strand cDNAs from various sources. Both the sense upstream primer and the antisense downstream primer were designed to have their 3' ends terminate within Cys residues.

The upstream sense primer encoded nucleotides 272-295 of SEQ ID NO:5 (region encoding amino acids 79-86); the downstream antisense primer encoded the complement of nucleotides 409-427 (region encoding amino acids 124-130). Standard PCR reactions were set up and run, using these primers and first strand cDNAs from various murine cell line or tissue sources. Thirty reaction cycles of 94°C for 30 seconds, 50°C for 30 seconds, and 72°C for 20 seconds were run. PCR products were anlyzed by electrophoresis, and specific bands were seen in several samples. The band from one sample was gel purified and DNA sequencing revealed that the sequence between the primers was approximately 85% identical to the corresponding human RANK nucleotide sequence.

A plasmid based cDNA library prepared from the murine fetal liver epithelium line FLE18 (one of the cell lines identified as positive in the PCR screen) was screened for full-length RANK cDNAs using murine RANK-specific oligonucleotide probes derived from the murine RANK sequence determined from sequencing the PCR product. Two cDNAs, one encoding the 5' end and one encoding the 3' end of full-length murine RANK (based on sequence comparison with the full-length human RANK) were recombined to generate a full-length murine RANK cDNA. The nucleotide and amino acid sequence of muRANK are shown in SEQ ID Nos:14 and 15.

The cDNA encodes a predicted Type 1 transmembrane protein having 625 amino acid residues, with a predicted 30 amino acid signal sequence, a 184 amino acid extracellular domain, a 21 amino acid transmembrane domain, and a 390 amino acid cytoplasmic tail. The extracellular region of muRANK displayed significant amino acid homology (69.7% identity, 80.8% similarity) to huRANK. Those of skill in the art will recognize that the actual cleavage site can be different from that predicted by computer, accordingly, the N-terminal of RANK may be from amino acid 25 to amino acid 35.

Other members of the TNF receptor superfamily have a region of amino acids between the transmembrane domain and the ligand binding domain that is referred to as a 'spacer' region, which is not necessary for ligand binding. In muRANK, the amino acids between 197 and 214 are predicted to form such a spacer region. Accordingly, a soluble form of RANK that terminates with an amino acid in this region is expected to retain the ability to bind a ligand for RANK in a specific manner. Preferred C-terminal amino acids for soluble RANK peptides are selected from the group consisting of amino acids 214, and 197 of SEQ ID NO:14, although other amino acids in the spacer region may be utilized as a C-terminus.

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#### **EXAMPLE 15**

This example illustrates the preparation of several different soluble forms of RANK and RANKL. Standard techniques of restriction enzyme cutting and ligation, in combination with PCR-based isolation of fragments for which no convenient restriction sites existed, were used. When PCR was utilized, PCR products were sequenced to ascertain whether any mutations had been introduced; no such mutations were found.

In addition to the huRANK/Fc described in Example 2, another RANK/Fc fusion protein was prepared by ligating DNA encoding amino acids 1-213 of SEQ ID NO:6, to DNA encoding amino acids 3-232 of the Fc mutein described previously (SEQ ID NO:8). A similar construct was prepared for murine RANK, ligating DNA encoding amino acids 1-213 of full-length murine RANK (SEQ ID NO:15) to DNA encoding amino acids 3-232 of the Fc mutein (SEQ ID NO:8).

A soluble, tagged, poly-His version of huRANKL was prepared by ligating DNA encoding the leader peptide from the immunoglobulin kappa chain (SEQ ID NO:16) to DNA encoding a short version of the FLAGTM tag (SEQ ID NO:17), followed by codons encoding Gly Ser, then a poly-His tag (SEQ ID NO:18), followed by codons encoding Gly Thr Ser, and DNA encoding amino acids 138-317 of SEQ ID NO:13. A soluble, poly-His tagged version of murine RANKL was prepared by ligating DNA encoding the CMV leader (SEQ ID NO:9) to codons encoding Arg Thr Ser, followed by DNA encoding poly-His (SEQ ID NO:18) followed by DNA encoding amino acids 119-294 of SEQ ID NO:11.

A soluble, oligomeric form of huRANKL was prepared by ligating DNA encoding the CMV leader (SEQ ID NO:9) to a codon encoding Asp followed by DNA ending a trimer-former "leucine" zipper (SEQ ID NO:19), then by codons encoding Thr Arg Ser followed by amino acids 138-317 of SEQ ID NO:13.

These and other constructs are prepared by routine experimentation. The various DNAs are then inserted into a suitable expression vector, and expressed. Particularly preferred expression vectors are those which can be used in mammalian cells. For example, pDC409 and pDC304, described herein, are useful for transient expression. For stable transfection, the use of CHO cells is preferred; several useful vectors are described in USSN 08/785,150, now allowed, for example, one of the 2A5-3 λ-derived expression vectors discussed therein.

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#### **EXAMPLE 16**

This example demonstrates that RANKL expression can be up-regulated on murine T cells. Cells were obtained from mesenteric lymph nodes of C57BL/6 mice, and activated with anti-CD3 coated plates, Concanavalin A (ConA) or phorbol myristate acetate in combination with ionomycin (anti-CD3: 500A2; Immunex Corporation, Seattle WA; ConA, PMA, ionomycin, Sigma, St. Louis, MO) substantially as described herein, and cultured from about 2 to 5 days. Expression of RANKL was evaluated in a three color analysis by FACS, using antibodies to the T cell markers CD4, CD8 and CD45RB, and RANK/Fc, prepared as described herein.

RANKL was not expressed on unstimulated murine T cells. T cells stimulated with either anti-CD3, ConA, or PMA/ionomycin, showed differential expression of RANKL: CD4\*/CD45RBLo and CD4\*/CD45RBHi cells were positive for RANKL, but CD8+ cells were not. RANKL was not observed on B cells, similar to results observed with human cells.

# EXAMPLE 17

This example illustrates the effects of murine RANKL on cell proliferation and activation. Various cells or cell lines representative of cells that play a role in an immune response (murine spleen, thymus and lymphnode) were evaluated by culturing them under conditions promoting their viability, in the presence or absence of RANKL. RANKL did not stimulate any of the tested cells to proliferate. One cell line, a macrophage cell line referred to as RAW 264.7 (ATCC accession number TIB 71) exhibited some signs of activation.

RAW cells constitutively produce small amounts of TNF- $\alpha$ . Incubation with either human or murine RANKL enhanced production of TNF- $\alpha$  by these cells in a dose

dependent manner. The results were not due to contamination of RANKL preparations with endotoxin, since boiling RANKL for 10 minutes abrogated TNF- $\alpha$  production, whereas a similar treatment of purified endotoxin (LPS) did not affect the ability of the LPS to stimulate TNF- $\alpha$  production. Despite the fact that RANKL activated the macrophage cell line RAW T64.7 for TNF- $\alpha$  production, neither human RANKL nor murine RANKL stimulated nitric oxide production by these cells.

# EXAMPLE 18

This example illustrates the effects of murine RANKL on growth and development of the thymus in fetal mice. Pregnant mice were injected with 1 mg of RANK/Fc or vehicle control protein (murine serum albumin; MSA) on days 13, 16 and 19 of gestation. After birth, the neonates continued to be injected with RANK/Fc intraperitoneally (IP) on a daily basis, beginning at a dose of 1 µg, and doubling the dose about every four days, for a final dosage of 4 µg. Neonates were taken at days 1, 8 and 15 post birth, their thymuses and spleens harvested and examined for size, cellularity and phenotypic composition.

A slight reduction in thymic size at day 1 was observed in the neonates born to the female injected with RANK/Fc; a similar decrease in size was not observed in the control neonates. At day 8, thymic size and cellularity were reduced by about 50% in the RANK/Fc-treated animals as compared to MSA treated mice. Phenotypic analysis demonstrated that the relative proportions of different T cell populations in the thymus were the same in the RANK/Fc mice as the control mice, indicating that the decreased cellularity was due to a global depression in the number of thymic T cells as opposed to a decrease in a specific population(s). The RANK/Fc-treated neonates were not significantly different from the control neonates at day 15 with respect to either size, cellularity or phenotype of thymic cells. No significant differences were observed in spleen size, cellularity or composition at any of the time points evaluated. The difference in cellularity on day 8 and not on day 15 may suggest that RANK/Fc may assert its effect early in thymic development.

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#### **EXAMPLE 19**

This example demonstrates that the C-terminal region of the cytoplasmic domain of RANK is important for binding of several different TRAF proteins. RANK contains at least two recognizable PXQX(X)T motifs that are likely TRAF docking sites. Accordingly, the importance of various regions of the cytoplasmic domain of RANK for TRAF binding was evaluated. A RANK/GST fusion protein was prepared substantially as described in Smith and Johnson, *Gene* 67:31 (1988), and used in the preparation of various truncations as described below.

Comparison of the nucleotide sequence of murine and human RANK indicated that there were several conserved regions that could be important for TRAF binding. Accordingly, a PCR-based technique was developed to facilitate preparation of various C-terminal truncations that would retain the conserved regions. PCR primers were designed to introduce a stop codon and restriction enzyme site at selected points, yielding the truncations described in Table 1 below. Sequencing confirmed that no undesired mutations had been introduced in the constructs.

Radio-labeled (35S-Met, Cys) TRAF proteins were prepared by in vitro translation using a commercially available reticulocyte lysate kit according to manufacturer's instructions (Promega). Truncated GST fusion proteins were purified substantially as described in Smith and Johnson (supra). Briefly, E. coli were transfected with an expression vector encoding a fusion protein, and induced to express the protein. The bacteria were lysed, insoluble material removed, and the fusion protein isolated by precipitation with glutathione-coated beads (Sepahrose 4B, Pharmacia, Uppsala Sweden)

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The beads were washed, and incubated with various radiolabeled TRAF proteins. After incubation and wash steps, the fusion protein/TRAF complexes were removed from the beads by boiling in 0.1% SDS + \(\beta\)-mercaptoethanol, and loaded onto 12% SDS gels (Novex). The gels were subjected to autoradiography, and the presence or absence of radiolabeled material recorded. The results are shown in Table 2 below.

Table 2: Binding of Various TRAF Proteins to the Cytoplasmic Domain of RANK

C terminal Truncations:	E206-S339	E206-Y421	E206-M476	E206-G544	Full length
TRAFI	-	•	-	-	++
TRAF2	•	-	•	-	++
TRAF3	•	-	-	-	++
TRAF4	-	•	-	-	•
TRAF5	-	-	-	-	+
TRAF6	-	+	+	+	++

These results indicate that TRAF1, TRAF2, TRAF3, TRAF 5 and TRAF6 bind to the most distal portion of the RANK cytoplasmic domain (between amino-acid G544 and A616). TRAF6 also has a binding site between S339 and Y421. In this experiment,

TRAF5 also bound the cytoplasmic domain of RANK.

WO 98/28426

#### PCT/US97/23775

#### SEQUENCE LISTING

- (1) GENERAL INFORMATION:
  - (i) APPLICANT: Immunex Corporation
  - (ii) TITLE OF INVENTION: Ligand for Receptor Activator of NF-kappaB
  - (iii) NUMBER OF SEQUENCES: 19
  - (iv) CORRESPONDENCE ADDRESS:
    - (A) ADDRESSEE: Immunex Corporation, Law Department
    - (B) STREET: 51 University Street
    - (C) CITY: Seattle
    - (D) STATE: WA
    - (E) COUNTRY: USA
    - (F) ZIP: 98101
  - (v) COMPUTER READABLE FORM:
    - (A) MEDIUM TYPE: Floppy disk

    - (B) COMPUTER: Apple Power Macintosh (C) OPERATING SYSTEM: Apple Operating System 7.5.5
    - (D) SOFTWARE: Microsoft Word for Power Macintosh 6.0.1
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- (viii) ATTORNEY/AGENT INFORMATION:
  (A) NAME: Perkins, Patricia Anne

  - (B) REGISTRATION NUMBER: 34,693
    (C) REFERENCE/DOCKET NUMBER: 2852-WO
  - (ix) TELECOMMUNICATION INFORMATION:
    (A) TELEPHONE: (206)587-0430

    - (B) TELEFAX: (206)233-0644
- (2) INFORMATION FOR SEQ ID NO:1:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 3115 base pairs
      (B) TYPE: nucleic acid

(C) STRANDEDNESS: single

			(D) :	POPO	LOGY:	: liı	near									
	(ii	í) M	DLECT	JLE :	TYPE:	cD!	AI									
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	(iv	/) AI	VTI-5	ENS	E: NO	)										
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	(vii	(	MEDI (A) I (B) C	IBR	RY:	BONE	-MAR	urow	DERI	VED	DENI	RITI	C CE	LLS		
	(ix	(	ATUR A) N B) L	IAME/				8								
	(xi	) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0:1:						
GCT	GCTG	CTG	CTCT	GCGC	GC T	GCTC	GCCC	G GC	TGCA	GTTI	TAT	CCAG	ААА	GAGC	TGTGTG	60
GAC	TCTC	TGC	CTGA	CCTC	AG T	GTTC	TTT	C AG	GTG	GCT	TTG	CAG	ATC	GCT	CCT	113
									Val 1	Ala	Leu	Gln	Ile 5	Ala	Pro	
CCA	TGT	ACC	AGT	GAG	AAG	CAT	TAT	GAG	CAT	CTG	GGA	CGG	TGC	TGT	AAC	161
	cys	10	ser	GIU	гÀЗ	His	171 15	Glu	His	Leu	Gly	Arg 20	Суз	Суѕ	Asn	
AAA	TGT	GAA	CCA	GGA	AAG	TAC	ATG	TCT	TCT	AAA	TGC	ACT	ACT	ACC	TCT	209
Lys	25	GIU	PIO	GIY	Lys	30	Met	Ser	Ser	Lys	Суs 35	Thr	Thr	Thr	Ser	
GAC	AGT	GTA	TGT	CTG	CCC	TGT	GGC	ccc	GAT	GAA	TAC	TTG	GAT	AGC	TGG	257
40	202	141	Cys	per	45	Сув	GIĀ	Pro	Asp	G1u 50	Tyr	Leu	Asp	Ser	Trp 55	
AAT Asn	GAA Glu	GAA	GAT	AAA	TGC	TTG	CTG	CAT	AAA	GTT	TGT	GAT Aśp	ACA	GGC	AAG	305
			11.50	60	Cys	Pañ	neu	nis	65 65	Val	Cys	Asp	Thr	Gly 70	Lys	
3CC Ala	CTG	GTG Val	GCC	GTG	GTC	GCC	GGC	AAC	AGC	ACG	ACC	ccc	CGG	CGC	TGC	353
			75	*41	<b>V</b> 41	ara	GIY	80	ser	Thr	Thr	Pro	Arg 85	Arg	Cys	
GCG	TGC	ACG	GCT	GGG	TAC	CAC	TGG	AGC	CAG	GAC	TGC	GAG	TGC	TGC	CGC	401
11 a	cys	90	ATA	GIY	Tyr	His	7xp 95	Ser	Gln	qaA	Сув	Glu 100	Cys	Суѕ	Arg	
GC	AAC	ACC	GAG	TGC	GCG	₽00	GGC	CTG	GGC	GCC	CAG	CAC	CCG	TTG	CAG	449
** A	105	rnr	GIU	суs	ΑΙΑ	Pro 110	Gly	Leu	Gly	Ala	Gln 115	His	Pro	Leu	Gln	
TC	AAC	AAG	GAC	ACA	GTG	TGC	AAA	CCT	TGC	СТТ	GCA	GGC	TAC	TTC	TCT	497
120		yy	กรม	THE	125	cys	гÀŝ	Pro	Суз	Leu 130	Ala	Gly	Tyr	Phe	Ser 135	

WO 98/28426 PCT/US97/23775 GAT GCC TTT TCC TCC ACG GAC AAA TGC AGA CCC TGG ACC AAC TGT ACC ASp Ala Phe Ser Ser Thr Asp Lys Cys Arg Pro Trp Thr Asp Cys Thr TTC CTT GGA AAG AGA GTA GAA CAT CAT GGG ACA GAG AAA TCC GAT GCG Phe Leu Gly Lys Arg Val Glu His His Gly Thr Glu Lys Ser Asp Ala 155 160 165593 GTT TGC AGT TCT TCT CTG CCA GCT AGA AAA CCA CCA AAT GAA CCC CAT Val Cys Ser Ser Leu Pro Ala Arg Lys Pro Pro Asn Glu Pro His GTT TAC TTG CCC GGT TTA ATA ATT CTG CTT CTC TTC GCG TCT GTG GCC Val Tyr Leu Pro Gly Leu Ile Ile Leu Leu Leu Pro Ala Ser Val Ala 689 CTG GTG GCT GCC ATC ATC TTT GGC GTT TGC TAT AGG AAA AAA GGG AAA Leu Val Ala Ala Ile Ile Phe Gly Val Cys Tyr Arg Lys Lys Gly Lys 200 210 215 GCA CTC ACA GCT AAT TTG TGG CAC TGG ATC AAT GAG GCT TGT GGC CGC Ala Leu Thr Ala Asn Leu Trp His Trp Ile Asn Glu Ala Cys Gly Arg 220 225 230 CTA AGT GGA GAT AAG GAG TCC TCA GGT GAC AGT TGT GTC AGT ACA CAC Leu Ser Gly Asp Lys Glu Ser Ser Gly Asp Ser Cys Val Ser Thr His 235 240 240 ACG GCA AAC TTT GGT CAG CAG GGA GCA TGT GAA GGT GTC TTA CTG CTG
Thr Ala Asn Phe Gly Gln Gln Gly Ala Cys Glu Gly Val Leu Leu
250 255 260 881 ACT CTG GAG GAG AAG ACA TTT CCA GAA GAT ATG TGC TAC CCA GAT CAA
Thr Leu Glu Glu Lys Thr Phe Pro Glu Asp Met Cys Tyr Pro Asp Gln
265 270 275 929 GGT GGT GTC TGT CAG GGC ACG TGT GTA GGA GGT GGT CCC TAC GCA CAA Gly Gly Val Cys Gln Gly Thr Cys Val Gly Gly Gly Pro Tyr Ala Gln 280 295 290 295 GGC GAA GAT GCC AGG ATG CTC TCA TTG GTC AGC AAG ACC GAG ATA GAG Gly Glu Asp Ala Arg Met Leu Ser Leu Val Ser Lys Thr Glu Ile Glu 1025 305 GAA GAC AGC TTC AGA CAG ATG CCC ACA GAA GAT GAA TAC ATG GAC AGG Glu Asp Ser Phe Arg Gln Met Pro Thr Glu Asp Glu Tyr Net Asp Arg 1073 CCC TCC CAG CCC ACA GAC CAG TTA CTG TTC CTC ACT GAG CCT GGA AGC 1121 Pro Ser Gin Pro Thr Asp Gin Leu Leu Phe Leu Thr Glu Pro Gly Ser AAA TCC ACA CCT CCT TTC TCT GAA CCC CTG GAG GTG GGG GAG AAT GAC Lys Ser Thr Pro Pro Phe Ser Glu Pro Leu Glu Val Gly Glu Asn Asp 345 AGT TTA AGC CAG TGC TTC ACG GGG ACA CAG AGC ACA GTG GGT TCA GAA Ser Leu Ser Gln Cys Phe Thr Gly Thr Gln Ser Thr Val Gly Ser Glu 1217 365

AGC TGC AAC TGC ACT GAG CCC CTG TGC AGG ACT GAT TGG ACT CCC ATG Ser Cys Asn Cys Thr Glu Pro Leu Cys Arg Thr Asp Trp Thr Pro Met 1265 TCC TCT GAA AAC TAC TTG CAA AAA GAG GTG GAC AGT GGC CAT TGC CCG Ser Ser Glu Asn Tyr Leu Gln Lys Glu Val Asp Ser Gly His Cys Pro 3951313 CAC TGG GCA GCC AGC CCC AGC CCC AAC TGG GCA GAT GTC TGC ACA GGC His Trp Ala Ala Ser Pro Ser Pro Asn Trp Ala Asp Val Cys Thr Gly 1361 TGC CGG AAC CCT CCT GGG GAG GAC TGT GAA CCC CTC GTG GGT TCC CCA Cys Arg Asn Pro Pro Gly Glu Asp Cys Glu Pro Leu Val Gly Ser Pro 1409 AAA CGT GGA CCC TTG CCC CAG TGC GCC TAT GGC ATG GGC CTT CCC CCT Lys Arg Gly Pro Leu Pro Gln Cys Ala Tyr Gly Met Gly Leu Pro Pro 1457 GAA GAA GAC AGC AGG ACG GAG GCC AGA GAC CAG CCC GAG GAT GGG Glu Glu Ala Ser Arg Thr Glu Ala Arg Asp Gln Pro Glu Asp Gly 1505 GCT GAT GGG AGG CTC CCA AGC TCA GCG AGG GCA GGT GCC GGG TCT GGA Ala Asp Gly Arg Leu Pro Ser Ser Ala Arg Ala Gly Ala Gly Ser Gly 475 480 485 1553 AGC TCC CCT GGT GGC CAG TCC CCT GCA TCT GGA AAT GTG ACT GGA AAC Ser Ser Pro Gly Gly Gln Ser Pro Ala Ser Gly Asn Val Thr Gly Asn 490 495 5001601 AGT AAC TCC ACG TTC ATC TCC AGG GGG CAG GTG ATG AAC TTC AAG GGC Ser Asn Ser Thr Phe Ile Ser Ser Gly Gln Val Met Asn Phe Lys Gly 1649 GAC ATC GTG GTC TAC GTC AGC CAG ACC TCG CAG GAG GGC GCG ASP 11e 11e Val Val Tyr Val Ser Gln Thr Ser Gln Glu Gly Ala Ala 520 535 1697 GCG GCT GCG GAG CCC ATG GGC CGC CCG GTG CAG GAG GAG ACC CTG GCG Ala Ala Ala Glu Pro Met Gly Arg Pro Val Glu Glu Thr Leu Ala CGC CGA GAC TCC TTC GCG GGG AAC GGC CCG CGC TTC CCG GAC CCG TGC Arg Arg Asp Ser Phe Ala Gly Asn Gly Pro Arg Phe Pro Asp Pro Cys 1793

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1888

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GGC GGC CCC GAG GGG CTG CGG GAG CCG GAG AAG GCC TCG AGG CCG GTG
Gly Gly Pro Glu Gly Leu Arg Glu Pro Glu Lys Ala Ser Arg Pro Val
570 575 580

CCCGAAGCTC GGAGCCAGGG CTCGCGAGGG CAGCACCGCA GCCTCTGCCC CAGCCCCGGC 1948
CACCCAGGGA TCGATCGGTA CAGTCGAGGA AGACCACCCG GCATTCTCTG CCCACTTTGC 2008
CTTCCAGGAA ATGGCTTTT CAGGAAGTGA ATTGATGAGG ACTGTCCCCA TGCCCACGGA 2068

CAG GAG CAA GGC GGG GCC AAG GCT TGA GCGCCCCCCA TGGCTGGGAG

Gln Glu Gln Gly Gly Ala Lys Ala 585 590

TGCTCAGCAG	CCCGCCGCAC	TGGGGCAGAT	GTCTCCCCTG	CCACTCCTCA	AACTCGCAGC	2128
agtaatttgt	GGCACTATGA	CAGCTATTTT	TATGACTATC	CTGTTCTGTG	GGGGGGGGT	2188
CTATGTTTTC	CCCCCATATT	TGTATTCCTT	TTCATAACTT	TTCTTGATAT	CTTTCCTCCC	2248
TCTTTTTTAA	TGTAAAGGTT	TTCTCAAAAA	TTCTCCTAAA	GGTGAGGGTC	TCTTTCTTTT	2308
CTCTTTTCCT	TTTTTTTTC	TTTTTTTGGC	AACCTGGCTC	TGGCCCAGGC	TAGAGTGCAG	2368
TGGTGCGATT	ATAGCCCGGT	GCAGCCTCTA	ACTCCTGGGC	TCAAGCAATC	CAAGTGATCC	2428
TCCCACCTCA	ACCTTCGGAG	TAGCTGGGAT	CACAGCTGCA	GGCCACGCCC	AGCTTCCTCC	2488
CCCCGACTCC	CCCCCCCAG	AGACACGGTC	CCACCATGTT	ACCCAGCCTG	GTCTCAAACT	2548
CCCCAGCTAA	AGCAGTCCTC	CAGCCTCGGC	CTCCCAAAGT	ACTGGGATTA	CAGGCGTGAG	2608
CCCCCACGCT	GGCCTGCTTT	ACGTATTTTC	TTTTGTGCCC	CTGCTCACAG	TGTTTTAGAG	2668
ATGGCTTTCC	CAGTGTGTGT	TCATTGTAAA	CACTTTTGGG	AAAGGGCTAA	ACATGTGAGG	2728
CCTGGAGATA	GTTGCTAAGT	TGCTAGGAAC	ATGTGGTGGG	ACTTTCATAT	TCTGAAAAAT	2788
GTTCTATATT	CTCATTTTTC	TAAAAGAAAG	AAAAAAGGAA	ACCCGATTTA	TTTCTCCTGA	2848
ATCTTTTAA	GTTTGTGTCG	TTCCTTAAGC	AGAACTAAGC	TCAGTATGTG	ACCTTACCCG	2908
CTAGGTGGTT	AATTTATCCA	TGCTGGCAGA	GGCACTCAGG	TACTTGGTAA	GCAAATTTCT	2968
AAAACTCCAA	GTTGCTGCAG	CTTGGCATTC	TTCTTATTCT	AGAGGTCTCT	CTGGAAAAGA	3028
TGGAGAAAAT	GAACAGGACA	TGGGGCTCCT	GGAAAGAAAG	GGCCCGGGAA	GTTCAAGGAA	3088
GAATAAAGTT	GAAATTTTAA	алалала				3115

### (2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 591 amino acids
   (B) TYPE: amino acid
   (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:
- Val Ala Leu Gln Ile Ala Pro Pro Cys Thr Ser Glu Lys His Tyr Glu 1 5 10 15
- His Leu Gly Arg Cys Cys Asn Lys Cys Glu Pro Gly Lys Tyr Met Ser 20 25 30
- Ser Lys Cys Thr Thr Thr Ser Asp Ser Val Cys Leu Pro Cys Gly Pro 35 40 45
- Asp Glu Tyr Leu Asp Ser Trp Asn Glu Glu Asp Lys Cys Leu Leu His  $50 \hspace{1.5cm} 55 \hspace{1.5cm} 60 \hspace{1.5cm}$

Lys Val Cys Asp Thr Gly Lys Ala Leu Val Ala Val Val Ala Gly Asn 65 70 75 80 Ser Thr Thr Pro Arg Arg Cys Ala Cys Thr Ala Gly Tyr His Trp Ser 85 90 95 Gln Asp Cys Glu Cys Cys Arg Arg Asn Thr Glu Cys Ala Pro Gly Leu 100 105 110 Gly Ala Gln His Pro Leu Gln Leu Asn Lys Asp Thr Val Cys Lys Pro 115 120 125 Cys Leu Ala Gly Tyr Phe Ser Asp Ala Phe Ser Ser Thr Asp Lys Cys 130 140 Arg Pro Trp Thr Asn Cys Thr Phe Leu Gly Lys Arg Val Glu His His 145 150 155 160 Gly Thr Glu Lys Ser Asp Ala Val Cys Ser Ser Ser Leu Pro Ala Arg 165 170 175 Lys Pro Pro Asn Glu Pro His Val Tyr Leu Pro Gly Leu Ile Ile Leu 180 185 190 Leu Leu Phe Ala Ser Val Ala Leu Val Ala Ala Ile Ile Phe Gly Val 195 200 205 Cys Tyr Arg Lys Lys Gly Lys Ala Leu Thr Ala Asn Leu Trp His Trp 210 215 220 Ile Asn Glu Ala Cys Gly Arg Leu Ser Gly Asp Lys Glu Ser Ser Gly 225 235 240 Asp Ser Cys Val Ser Thr His Thr Ala Asn Phe Gly Gln Gln Gly Ala 245 250 255 Cys Glu Gly Val Leu Leu Leu Thr Leu Glu Glu Lys Thr Phe Pro Glu 260 265 270 Asp Met Cys Tyr Pro Asp Gln Gly Gly Val Cys Gln Gly Thr Cys Val 275 280 285 Gly Gly Gly Pro Tyr Ala Gln Gly Glu Asp Ala Arg Met Leu Ser Leu 290 295 300 Val Ser Lys Thr Glu Ile Glu Glu Asp Ser Phe Arg Gln Met Pro Thr 305 310 315 320 Glu Asp Glu Tyr Met Asp Arg Pro Ser Gln Pro Thr Asp Gln Leu Leu 325 330 335 Phe Leu Thr Glu Pro Gly Ser Lys Ser Thr Pro Pro Phe Ser Glu Pro 340 345 350 Leu Glu Val Gly Glu Asn Asp Ser Leu Ser Gln Cys Phe Thr Gly Thr 355 360 365 Gln Ser Thr Val Gly Ser Glu Ser Cys Asn Cys Thr Glu Pro Leu Cys 370 380

Arg Thr Asp Trp Thr Pro Met Ser Ser Glu Asn Tyr Leu Gln Lys Glu

Val Asp Ser Gly His Cys Pro His Trp Ala Ala Ser Pro Ser Pro Asn 405 410 415

Trp Ala Asp Val Cys Thr Gly Cys Arg Asn Pro Pro Gly Glu Asp Cys
420 425 430

Glu Pro Leu Val Gly Ser Pro Lys Arg Gly Pro Leu Pro Gln Cys Ala 435 440 445

Tyr Gly Met Gly Leu Pro Pro Glu Glu Glu Ala Ser Arg Thr Glu Ala 450 460

Arg Asp Gln Pro Glu Asp Gly Ala Asp Gly Arg Leu Pro Ser Ser Ala 465 470 475 480

Arg Ala Gly Ala Gly Ser Gly Ser Ser Pro Gly Gly Gln Ser Pro Ala
485 490 495

Ser Gly Asn Val Thr Gly Asn Ser Asn Ser Thr Phe Ile Ser Ser Gly 500 505 510

Gln Val Met Asn Phe Lys Gly Asp Ile Ile Val Val Tyr Val Ser Gln 515 520 525

Thr Ser Gln Glu Gly Ala Ala Ala Ala Ala Glu Pro Met Gly Arg Pro 530 535 540

Val Gln Glu Glu Thr Leu Ala Arg Arg Asp Ser Phe Ala Gly Asn Gly 545 550 555 560

Pro Arg Phe Pro Asp Pro Cys Gly Gly Pro Glu Gly Leu Arg Glu Pro 565 570 575

Glu Lys Ala Ser Arg Pro Val Gln Glu Gln Gly Gly Ala Lys Ala 580 585 590

#### (2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1391 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: HOMO SAPIENS
- (vii) IMMEDIATE SOURCE:
  - (A) LIBRARY: BONE-MARROW DERIVED DENDRITIC CELLS
  - (B) CLONE: 9D-15C

#### PCT/US97/23775

(ix)	PEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 39..1391

#### (xi) SEQUENCE DESCRIPTION: SEO ID NO:3

	(X1	) SE	QUEN	CED	BSCR	IPTI	ON:	SEQ	ID N	0:3:						
cca	CTGA	.GGC	cccc	GCGC	CC G	CCAG	CCTG	ጥ ሮሮ	CGCG				CG C			53
AIG	Arg	Arg	Arg	10		Phe	Ala	Leu	Leu 15	Leu	Leu	Суѕ	Ala	Leu 20	Leu	101
Ala	Arg	Leu	25	Val	GCT Ala	Leu	Gln	Ile 30	Ala	Pro	Pro	Суз	Thr 35	Ser	Glu	149
гÀЗ	пть	40	GIU	H1S	CTG Leu	Gly	Arg 45	Сла	Cys	Asn	Lys	Cys 50	Glu	Pro	Gly	197
гуу	55	mec	ser	Ser	AAA Lys	60 Cys	Thr	Thr	Thr	Ser	Asp 65	Ser	Val	Cys	Leu	245
70	cys	GIY	Pro	Asp	GAA Glu 75	Tyr	Leu	qeA	Ser	Trp 80	Asn	Glu	Glu	Asp	Lys 85	293
Cys	Leu	rea	HIS	90 50	GTT Val	Суз	Asp	Thr	95 95	Lys	Ala	Leu	Val	Ala 100	Val	341
Val	VIG	GIĀ	105	ser	ACG Thr	Thr	Pro	Arg 110	Arg	Суs	Ala	Cys	Thr 115	Ala	Gly	389
тут	нıs	120	Ser	Gln	GAC Asp	СЛЗ	Glu 125	Суз	Cys	Arg	Arg	Asn 130	Thr	Glu	Cys	437
MIA	135	СІУ	Leu	Gly	GCC Ala	Gln 140	His	Pro	Leu	Gln	Leu 145	Asn	Lys	Asp	Thr	485
150	Cys	Lys	Pro	Сув	CTT Leu 155	Ala	Gly	Tyr	Phe	Ser 160	Asp	Ala	Phe	Ser	Ser 165	533
Ini	ASP	гÀв	Cys	170	CCC Pro	Trp	Thr	Asn	Cys 175	Thr	Phe	Leu	Gly	Lys 180	Arg	581
GTA Val	GAA Glu	CAT His	CAT His 185	GGG Gly	ACA Thr	GAG Glu	AAA Lys	TCC Ser 190	GAT Asp	GCG Ala	GTT Val	TGC Cys	AGT Ser 195	TCT Ser	TCT Ser	629

CTG Lev	CCA Pro	GCT Ala 200	AGA Arg	AAA Lys	CCA Pro	CCA Pro	AAT Asn 205	Glu	CCC Pro	CAT His	GTT Val	TAC Tyr 210	TTG	CCC	GGT Gly	677
TTA Leu	Ile 215	TIE	CTG Leu	CTT	CTC Leu	TTC Phe 220	Ala	TCT	GTG Val	GCC Ala	CTG Leu 225	GTG Val	GCT Ala	GCC Ala	ATC Ile	725
ATC Ile 230	rne	GGC	GTT Val	TGC Cys	TAT Tyr 235	AGG Arg	AAA Lys	AAA Lys	GGG Gly	AAA Lys 240	Ala	CTC Leu	ACA Thr	GCT Ala	AAT Asn 245	773
TTG Leu	TGG Trp	CAC His	TGG Trp	ATC 11e 250	AAT Asn	GAG Glu	GCT Ala	TGT Cys	GGC Gly 255	CGC Arg	CTA Leu	AGT Ser	GGA Gly	GAT Asp 260	AAG Lys	821
GAG Glu	TCC Ser	TCA Ser	GGT Gly 265	GAC Asp	AGT Ser	TGT Cys	GTC Val	AGT Ser 270	ACA Thr	CAC His	ACG Thr	GCA Ala	AAC Asn 275	TTT Phe	GGT Gly	869
CAG Gln	CAG Gln	GGA Gly 280	GCA Ala	TGT Cys	GAA Glu	GGT Gly	GTC Val 285	TTA Leu	CTG Leu	CTG Leu	ACT Thr	CTG Leu 290	GAG Glu	GAG Glu	AAG Lys	917
ACA Thr	TTT Phe 295	CCA Pro	GAA Glu	GAT Asp	ATG Met	TGC Cys 300	TAC Tyr	CCA Pro	GAT Asp	CAA Gln	GGT Gly 305	GGT Gly	GTC Val	TGT Cys	CAG Gln	965
GGC Gly 310	ACG Thr	TGT Cys	GTA Val	GGA Gly	GGT Gly 315	GGT Gly	CCC Pro	TAC Tyr	GCA Ala	CAA Gln 320	GGC Gly	GAA Glu	GAT Asp	GCC Ala	AGG Arg 325	013
ATG Met	CTC Leu	TCA Ser	TTG Leu	GTC Val 330	AGC Ser	AAG Lys	ACC Thr	GAG Glu	ATA Ile 335	GAG Glu	GAA Glu	GAC Asp	AGC Ser	TTC Phe 340	AGA Arg	061
CAG Gln	ATG Met	CCC Pro	ACA Thr 345	GAA Glu	GAT Asp	GAA Glu	TAC Tyr	ATG Met 350	gac Asp	AGG Arg	CCC Pro	TCC Ser	CAG Gln 355	CCC Pro	ACA Thr	109
GAC Asp	CAG Gln	TTA Leu 360	CTG Leu	TTC Phe	CTC Leu	ACT Thr	GAG Glu 365	CCT Pro	GGA Gly	AGC Ser	AAA Lys	TCC Ser 370	ACA Thr	CCT Pro	CCT Pro	157
TTC Phe	TCT Ser 375	GAA Glu	CCC Pro	CTG Leu	GAG Glu	GTG Val 380	GJ Y GGG	GAG Glu	AAT Asn	GAC Asp	AGT Ser 385	TTA Leu	AGC Ser	CAG Gln	TGC Cys	205
TTC Phe 390	ACG Thr	CJA CCC	ACA Thr	CAG Gln	AGC Ser 395	ACA Thr	GTG Val	GGT Gly	TCA Ser	GAA Glu 400	AGC Ser	TGC Cys	AAC Asn	TGC Cys	ACT Thr 405	253
GAG Glu	CCC Pro	CTG Leu	TGC Cys	AGG Arg 410	ACT Thr	GAT Asp	TGG Trp	ACT Thr	CCC Pro 415	ATG Met	TCC Ser	TCT Ser	GAA Glu	AAC Asn 420	TAC Tyr	301
TTG Leu	CAA Gln	AAA Lys	GAG Glu 425	GTG Val	GAC Asp	AGT Ser	Gly	CAT His 430	TGC Cys	CCG Pro	CAC His	TGG Trp	GCA Ala 435	GCC Ala	AGC Ser	349

CCC AGC CCC AAC TGG GCA GAT GTC TGC ACA GGC TGC CGG AAC Pro Ser Pro Asn Trp Ala Asp Val Cys Thr Gly Cys Arg Asn 445

### (2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 451 amino acids
    (B) TYPE: amino acid

  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Ala Pro Arg Ala Arg Arg Arg Pro Leu Phe Ala Leu Leu 1 5 10 15

Leu Cys Ala Leu Leu Ala Arg Leu Gln Val Ala Leu Gln Ile Ala Pro 20 25 30

Pro Cys Thr Ser Glu Lys His Tyr Glu His Leu Gly Arg Cys Cys Asn 35 40 45

Lys Cys Glu Pro Gly Lys Tyr Met Ser Ser Lys Cys Thr Thr Thr Ser 50 60

Asp Ser Val Cys Leu Pro Cys Gly Pro Asp Glu Tyr Leu Asp Ser Trp 65 70 75 80

Asn Glu Glu Asp Lys Cys Leu Leu His Lys Val Cys Asp Thr Gly Lys 85 90 95

Ala Leu Val Ala Val Val Ala Gly Asn Ser Thr Thr Pro Arg Arg Cys

Ala Cys Thr Ala Gly Tyr His Trp Ser Gln Asp Cys Glu Cys Cys Arg 115 120 125

Arg Asn Thr Glu Cys Ala Pro Gly Leu Gly Ala Gln His Pro Leu Gln 130 135 140

Leu Asn Lys Asp Thr Val Cys Lys Pro Cys Leu Ala Gly Tyr Phe Ser 145 150 155 160

Asp Ala Phe Ser Ser Thr Asp Lys Cys Arg Pro Trp Thr Asn Cys Thr 165 170 175

Phe Leu Gly Lys Arg Val Glu His His Gly Thr Glu Lys Ser Asp Ala 180 185 190

Val Cys Ser Ser Ser Leu Pro Ala Arg Lys Pro Pro Asn Glu Pro His 195 200 205

Val Tyr Leu Pro Gly Leu Ile Ile Leu Leu Leu Phe Ala Ser Val Ala 210 215 220

Leu Val Ala Ala Ile Ile Phe Gly Val Cys Tyr Arg Lys Lys Gly Lys 225 230 235 240

Ala Leu Thr Ala Asn Leu Trp His Trp Ile Asn Glu Ala Cys Gly Arg 245 250 255

Leu Ser Gly Asp Lys Glu Ser Ser Gly Asp Ser Cys Val Ser Thr His 260 265 270

Thr Ala Asn Phe Gly Gln Gln Gly Ala Cys Glu Gly Val Leu Leu 275 280 285

Thr Leu Glu Glu Lys Thr Phe Pro Glu Asp Met Cys Tyr Pro Asp Gln 290 295 300

Gly Gly Val Cys Gln Gly Thr Cys Val Gly Gly Gly Pro Tyr Ala Gln 305 310 315 320

Gly Glu Asp Ala Arg Met Leu Ser Leu Val Ser Lys Thr Glu Ile Glu 325 330 335

Glu Asp Ser Phe Arg Gln Met Pro Thr Glu Asp Glu Tyr Met Asp Arg 340 345 350

Pro Ser Gln Pro Thr Asp Gln Leu Leu Phe Leu Thr Glu Pro Gly Ser 355 360 365

Lys Ser Thr Pro Pro Phe Ser Glu Pro Leu Glu Val Gly Glu Asn Asp 370 375 380

Ser Leu Ser Gln Cys Phe Thr Gly Thr Gln Ser Thr Val Gly Ser Glu 385 390 395 400

Ser Cys Asn Cys Thr Glu Pro Leu Cys Arg Thr Asp Trp Thr Pro Met
405 410 415

Ser Ser Glu Asn Tyr Leu Gln Lys Glu Val Asp Ser Gly His Cys Pro
420 425 430

His Trp Ala Ala Ser Pro Ser Pro Asn Trp Ala Asp Val Cys Thr Gly
435 445

Cys Arg Asn 450

- (2) INFORMATION FOR SEQ ID NO:5:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 3136 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (vi) ORIGINAL SOURCE:
    (A) ORGANISM: HOMO SAPIENS
  - (vii) IMMEDIATE SOURCE:

(A) LIBRARY: BONE-MARROW DERIVED DENDRITIC CELLS (B) CLONE: FULL LENGTH RANK

(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 39..1886

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

				GGCG						r	let i	Ala	Pro .	Arg	Ala 5	53
			_	10	)		. ATG	ь	1 15	Leu	ı Leı	и Су:	a Ala	1 Lei 20	-	101
			25	5		, Den	G11	30	Ala	Pro	Pro	о Суа	Th:	Ser	GAG Glu	149
_		40	)		Deu	GLY	45	cys	Cys	Asn	Lys	Cys 50	Glu	Pro	GGA Gly	197
AAC Lys	TAC Tyr 55	Met	S TCT	TCT Ser	AAA Lys	TGC Cys 60	ACT Thr	ACT	ACC	TCT	GAC Asp 65	Ser	GTA Val	TG1 Cys	CTG	245
70		•			75	ıyı	reu	Asp	Ser	Trp 80	naA	Glu	Glu	Asp	85	293
-				AAA Eys 00	•41	cys	АБР	inr	95	Lys	Ala	Leu	Val	Ala 100	Val	341
GTC Val	GCC Ala	GGC	AAC Asn 105	AGC Ser	ACG Thr	ACC Thr	CCC Pro	CGG Arg 110	CGC Arg	TGC Cys	GCG Ala	TGC Cys	ACG Thr 115	GCT Ala	GGG Gly	389
		120		CAG Gln	op	cys	125	cys	Cys	Arg	Arg	Asn 130	Thr	Glu	Cys	437
GCG Ala	CCG Pro 135	GGC Gly	CTG Leu	GGC GIy	GCC Ala	CAG Gln 140	CAC His	CCG Pro	TTG Leu	CAG Gln	CTC Leu 145	AAC Asn	AAG Lys	GAC Asp	ACA Thr	485
GTG Val 150	TGC Cys	AAA Lys	CCT Pro	TGC Cys	CTT Leu 155	GCA Ala	GGC Gly	TAC Tyr	TTC Phe	TCT Ser 160	Gat Asp	GCC Ala	TTT Phe	TCC Ser	TCC Ser 165	533
ACG Thr	GAC Asp	AAA Lys	TGC Cys	AGA Arg 170	CCC Pro	TGG Trp	ACC Thr	AAC Asn	TGT Cys 175	ACC Thr	TTC Phe	CTT Leu	GGA Gly	AAG Lys 180	AGA Arg	581
GTA Val	GAA Glu	CAT His	CAT His 185	GCG Gly	ACA Thr	GAG Glu	rys .	TCC Ser 190	GAT Asp	GCG Ala	GTT Val	TGC Cys	AGT Ser 195	TCT Ser	TCT Ser	629

CTG	CCA	GCT	AGA	AAA	CCA	CCA	AAT	GAA	ccc	CAT	Calab	<b>ም</b> እ ^	TI-TI-C	ccc	~~	
Leu	Pro	200	Arg	Lys	Pro	Pro	Asn 205	Glu	Pro	His	Val	Туг 210	Leu	Pro	Gly	677
TTA Leu	ATA Ile 215	ATT	CTG Leu	CTT	CTC Leu	TTC Phe 220	GCG Ala	TCT Ser	GTG Val	GCC Ala	CTG Leu 225	GTG Val	GCT Ala	GCC Ala	ATC Ile	725
ATC Ile 230	TTT	GGC Gly	GTT Val	TGC Cys	ТАТ Тут 235	AGG Arg	AAA Lys	AAA Lys	GGG Gly	AAA Lys 240	GCA Ala	CTC Leu	ACA Thr	GCT Ala	AAT Asn 245	773
TTG Leu	TGG Trp	CAC His	TGG Trp	ATC Ile 250	AAT Asn	GAG Glu	GCT Ala	TGT Cys	GGC Gly 255	CGC Arg	CTA Leu	AGT Ser	GGA Gly	GAT Asp 260	AAG Lys	821
GAG Glu	TCC Ser	TCA Ser	GGT Gly 265	GAC Asp	AGT Ser	TGT Cys	GTC Val	AGT Ser 270	ACA Thr	CAC His	ACG Thr	GCA Ala	AAC Asn 275	TTT Phe	GGT Gly	869
CAG Gln	CAG Gln	GGA Gly 280	GCA Ala	TGT Cys	GAA Glu	GGT Gly	GTC Val 285	TTA Leu	CTG Leu	CTG Leu	ACT Thr	CTG Leu 290	GAG Glu	GAG Glu	AAG Lys	917
ACA Thr	TTT Phe 295	CCA Pro	GAA Glu	GAT Asp	ATG Met	TGC Cys 300	TAC Tyr	CCA Pro	gat Asp	CAA Gln	GGT Gly 305	GGT Gly	GTC Val	TGT Cys	CAG Gln	965
GGC Gly 310	ACG Thr	TGT Cys	GTA Val	GGA Gly	GGT Gly 315	GGT Gly	CCC Pro	TAC Tyr	GCA Ala	CAA Gln 320	GGC Gly	GAA Glu	GAT Asp	GCC Ala	AGG Arg 325	1013
ATG Met	CTC Leu	TCA Ser	TTG Leu	GTC Val 330	AGC Ser	AAG Lys	ACC Thr	Glu	ATA Ile 335	GAG Glu	GAA Glu	GAC Asp	AGC Ser	TTC Phe 340	AGA Arg	1061
CAG Gln	ATG Met	CCC Pro	ACA Thr 345	GAA Glu	GAT Asp	GAA Glu	TAC Tyr	ATG Met 350	Asp	AGG Arg	CCC Pro	TCC Ser	CAG Gln 355	CCC Pro	ACA Thr	1109
GAC Asp	CAG Gln	TTA Leu 360	CTG Leu	TTC Phe	CTC Leu	ACT Thr	GAG Glu 365	CCT Pro	GGA Gly	AGC Ser	AAA Lys	TCC Ser 370	ACA Thr	CCT Pro	CCT Pro	1157
TTC Phe	TCT Ser 375	GAA Glu	CCC Pro	CTG Leu	GAG Glu	GTG Val 380	GGG Gly	GAG Glu	AAT Asn	GAC Asp	AGT Ser 385	TTA Leu	AGC Ser	CAG Gln	TGC Cys	1205
TTC Phe 390	ACG Thr	GGG Gly	ACA Thr	CAG Gln	AGC Ser 395	ACA Thr	GTG Val	GGT Gly	TCA Ser	GAA Glu 400	AGC Ser	TGC Cys	AAC Asn	TGC Cys	ACT Thr 405	1253
GAG Glu	CCC Pro	CTG Leu	TGC Cys	AGG Arg 410	ACT Thr	GAT Asp	TGG Trp	ACT Thr	CCC Pro 415	ATG Met	TCC Ser	TCT Ser	GAA Glu	AAC Asn 420	TAC Tyr	1301
TTG Leu	CAA Gln	гÀв	GAG Glu 425	GTG Val	GAC Asp	AGT Ser	GGC Gly	CAT His 430	TGC Cys	CCG Pro	CAC His	TGG Trp	GCA Ala 435	GCC Ala	AGC Ser	1349

CCC Pro	AGC Ser	CCC Pro 440	AAC Asn	TGG Trp	GCA Ala	GAT Asp	GTC Val 445	TGC Cys	ACA Thr	GGC Gly	TCC ayo	CGG Arg 450	AAC Asn	CCT Pro	CCT Pro	1397
GGG Gly	GAG Glu 455	GAC Asp	TGT Cys	GAA Glu	CCC Pro	CTC Leu 460	GTG Val	GGT Gly	TCC Ser	CCA Pro	AAA Lys 465	CGT Arg	GGA Gly	CCC Pro	TTG Leu	1445
CCC Pro 470	CAG Gln	TGC Cys	GCC Ala	TAT Tyr	GGC Gly 475	ATG Met	GGC Gly	CTT Leu	CCC Pro	CCT Pro 480	GAA Glu	GAA Glu	GAA Glu	GCC Ala	AGC Ser 485	1493
AGG Arg	ACG Thr	GAG Glu	GCC Ala	AGA Arg 490	GAC Asp	CAG Gln	CCC Pro	GAG Glu	GAT Asp 495	GGG Gly	GCT Ala	GAT Asp	G13 G13	AGG Arg 500	CTC Leu	1541
CCA Pro	AGC Ser	TCA Ser	GCG Ala 505	AGG Arg	GCA Ala	GCT Gly	GCC Ala	GGG Gly 510	TCT Ser	GGA Gly	AGC Ser	TCC Ser	CCT Pro 515	GGT Gly	GJA GGC	1589
CAG Gln	TCC Ser	CCT Pro 520	GCA Ala	TCT Ser	GGA Gly	AAT Asn	GTG Val 525	ACT Thr	GGA Gly	AAC Asn	AGT Ser	AAC Asn 530	TCC Ser	ACG Thr	TTC Phe	1637
ATC Ile	TCC Ser 535	AGC Ser	GGG Gly	CAG Gln	GTG Val	ATG Met 540	AAC Asn	TTC Phe	AAG Lys	GGC Gly	GAC Asp 545	ATC Ile	ATC Ile	GTG Val	GTC Val	1685
TAC Tyr 550	GTC Val	AGC Ser	CAG Gln	ACC Thr	TCG Ser 555	CAG Gln	GAG Glu	GGC Gly	GCG Ala	GCG Ala 560	GCG Ala	GCT Ala	GCG Ala	GAG Glu	CCC Pro 565	1733
ATG Met	GGC Gly	CGC Arg	CCG Pro	GTG Val 570	CAG Gln	GAG Glu	GAG. Glu	ACC Thr	CTG Leu 575	GCG Ala	CGC Arg	CGA Arg	GAC Asp	TCC Ser 580	TTC Phe	1781
GCG Ala	GGG Gly	AAC Asn	GGC Gly 585	CCG Pro	CGC Arg	TTC Phe	CCG Pro	GAC Asp 590	CCG Pro	TGC Cys	GGC Gly	GGC Gly	CCC Pro 595	GAG Glu	GGG Gly	1829
CTG Leu	CGG Arg	GAG Glu 600	CCG Pro	GAG Glu	AAG Lys	GCC Ala	TCG Ser 605	ACG Arg	CCG Pro	GTG Val	CAG Gln	GAG Glu 610	CAA Gln	GGC Gly	GGG Gly	1877
GCC Ala	AAG Lys 615	GCT Ala	TGAC	CGCC	ec c	CATO	GCT	G G	GCCC	GAAG	CTC	GGAC	CCA			1926
GGGC	TCGC	GA C	GGC!	GCAC	c GC	AGCC	тстс	.ccc	CAGO	ccc	GGC	ACC	AG (	GATO	GATCG	1986
GTAC	AGTO	GA G	GAAC	ACCA	c co	GGC?	TTCI	CTC	CCC#	CTT	TGCC	TTC	AG (	GAAA1	GGGCT	2046
TTTC	AGGA	AG 1	GAAT	TGAT	G AG	GACT	GTC	CCZ	TGCC	CAC	GGA7	CTC	AG (	CAGCO	cecce	2106
CACI	GGGG	CA G	atg:	CTC	C CI	GCC	CTCC	TC	LAACI	rege	AGC!	GTA	TT	rgrgg	CACTA	2166
TGAC	AGCT	r ta	TTT	TGAC	ra Te	CCTC	TTCT	GTC	GGGG	GGG	CCTC	TATO	TT 1	TCCC	CCCAT	2226
ATTI	GTAI	TC C	TTT.	CATA	LA CI	TTT	TTGA	TAT	CTT	CCT	CCC1	CTT	TT :	TAATO	STAAAG	2286
GITI	TCTC	AA A	AATT	CTCC	T AA	AGG1	GAGG	GTY	TCTT	TCT	TTT	TCT	TT (	ccTT1	TTTTT	2346

TTCTTTTTTT GGCAACCTGG CTCTGGCCCA GGCTAGAGTG CAGTGGTGCG ATTATAGCCC 2406
GGTGCAGCCT CTAACTCCTG GGCTCAAGCA ATCCAAGTGA TCCTCCCACC TCAACCTTCG 2466
GAGTAGCTGG GATCACAGCT GCAGGCCACG CCCAGCTTCC TCCCCCGCC TCCCCCCCC 2526
CAGAGACACG GTCCCACCAT GTTACCCAGC CTGGTCTCAA ACTCCCCAGC TAAAGCAGTC 2586
CTCCAGCCTC GGCCTCCCAA AGTACTGGGA TTACAGGCGT GAGCCCCCAC GCTGGCCTGC 2646
TTTACGTATT TTCTTTTGTG CCCCTGCTCA CAGTGTTTTA GAGATGGCTT TCCCAGTGTG 2706
AGTTGCTAGG AACACTTTT GGGAAAGGGC TAAACATGTG AGGCCTGGAG ATAGTTGCTA 2766
AGTTGCTAGG AACACTTTT GGGAAAGGGC TAAACATGTG AGGCCTGGAG ATAGTTGCTA 2826
TTCTAAAAAAAAAA AAGAAAAAAA GAAACCCGAT TTATTCTCC TGAATCTTT TAAGTTTTGT 2886
TCGTTCCTTA AGCAGAACTA AGCTCAGTAT GTGACCTTAC CCGCTAGGTG GTTAATTTAT 2946
CCATGCTGGC AGAGGCACTC AGGTACTTGG TAAGCAAATT TCTAAAACCC CAAGTTGCTG 3006
CAGCTTGGCA TTCTTCTTAT TCTAGAGGTC TCTCTGGAAA AGATGAGAAA AATGAACAGG 3066
ACATGGGGCT CCTGGAAAGA AAGGGCCCGG GAAGTTCAAG GAAGAATAAA GTTGAAATTT 3126
TAAAAAAAAAA

- (2) INFORMATION FOR SEQ ID NO:6:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 616 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:
- Met Ala Pro Arg Ala Arg Arg Arg Pro Leu Phe Ala Leu Leu Leu 1 5 10 15
- Leu Cys Ala Leu Leu Ala Arg Leu Gln Val Ala Leu Gln Ile Ala Pro 20 25 30
- Pro Cys Thr Ser Glu Lys His Tyr Glu His Leu Gly Arg Cys Cys Asn 35 40 45
- Lys Cys Glu Pro Gly Lys Tyr Met Ser Ser Lys Cys Thr Thr Thr Ser 50 55 60
- Asp Ser Val Cys Leu Pro Cys Gly Pro Asp Glu Tyr Leu Asp Ser Trp 65 70 75 80
- Asn Glu Glu Asp Lys Cys Leu Leu His Lys Val Cys Asp Thr Gly Lys  $85 \hspace{1cm} 90 \hspace{1cm} 95$
- Ala Leu Val Ala Val Val Ala Gly Asn Ser Thr Thr Pro Arg Arg Cys 100  $\phantom{000}105\phantom{000}$  110

Ala Cys Thr Ala Gly Tyr His Trp Ser Gln Asp Cys Glu Cys Cys Arg 115 120 125 Arg Asn Thr Glu Cys Ala Pro Gly Leu Gly Ala Gln His Pro Leu Gln 130 135 140 Leu Asn Lys Asp Thr Val Cys Lys Pro Cys Leu Ala Gly Tyr Phe Ser 145 150 155 160 Asp Ala Phe Ser Ser Thr Asp Lys Cys Arg Pro Trp Thr Asn Cys Thr 165 170 175 Phe Leu Gly Lys Arg Val Glu His His Gly Thr Glu Lys Ser Asp Ala 180 185 190 Val Cys Ser Ser Ser Leu Pro Ala Arg Lys Pro Pro Asn Glu Pro His 195 200 205 Val Tyr Leu Pro Gly Leu Ile Ile Leu Leu Leu Phe Ala Ser Val Ala 210 215 220 Leu Val Ala Ala Ile Ile Phe Gly Val Cys Tyr Arg Lys Lys Gly Lys 225 230 235 240 Ala Leu Thr Ala Asn Leu Trp His Trp Ile Asn Glu Ala Cys Gly Arg 245 250 255 Leu Ser Gly Asp Lys Glu Ser Ser Gly Asp Ser Cys Val Ser Thr His 260 265 270 Thr Ala Asn Phe Gly Gln Gln Gly Ala Cys Glu Gly Val Leu Leu 275 280 285 Thr Leu Glu Glu Lys Thr Phe Pro Glu Asp Met Cys Tyr Pro Asp Gln 290 295 300 Gly Gly Val Cys Gln Gly Thr Cys Val Gly Gly Gly Pro Tyr Ala Gln 305 310 315 320Gly Glu Asp Ala Arg Met Leu Ser Leu Val Ser Lys Thr Glu Ile Glu 325 330 335 Glu Asp Ser Phe Arg Gln Met Pro Thr Glu Asp Glu Tyr Met Asp Arg 340 345 350 Pro Ser Gln Pro Thr Asp Gln Leu Leu Phe Leu Thr Glu Pro Gly Ser 355 360 365Lys Ser Thr Pro Pro Phe Ser Glu Pro Leu Glu Val Gly Glu Asn Asp 370 375 380 Ser Leu Ser Gln Cys Phe Thr Gly Thr Gln Ser Thr Val Gly Ser Glu 385 390 395 400 Ser Cys Asn Cys Thr Glu Pro Leu Cys Arg Thr Asp Trp Thr Pro Met 405 410 415 Ser Ser Glu Asn Tyr Leu Gln Lys Glu Val Asp Ser Gly His Cys Pro 420 425 430

His Trp Ala Ala Ser Pro Ser Pro Asn Trp Ala Asp Val Cys Thr Gly 435 440 445

Cys Arg Asn Pro Pro Gly Glu Asp Cys Glu Pro Leu Val Gly Ser Pro
450 455 460

Lys Arg Gly Pro Leu Pro Gln Cys Ala Tyr Gly Met Gly Leu Pro Pro 480

Glu Glu Glu Ala Ser Arg Thr Glu Ala Arg Asp Gln Pro Glu Asp Gly 485 490 495

Ala Asp Gly Arg Leu Pro Ser Ser Ala Arg Ala Gly Ala Gly Ser Gly 500 505 510

Ser Ser Pro Gly Gly Gln Ser Pro Ala Ser Gly Asn Val Thr Gly Asn 515 520 525

Ser Asn Ser Thr Phe Ile Ser Ser Gly Gln Val Met Asn Phe Lys Gly 530 535 540

Asp Ile Ile Val Val Tyr Val Ser Gln Thr Ser Gln Glu Gly Ala Ala 545 550 550 560

Ala Ala Ala Glu Pro Met Gly Arg Pro Val Gln Glu Glu Thr Leu Ala 565 570 575

Arg Arg Asp Ser Phe Ala Gly Asn Gly Pro Arg Phe Pro Asp Pro Cys 580 585 590

Gly Gly Pro Glu Gly Leu Arg Glu Pro Glu Lys Ala Ser Arg Pro Val 595 600 605

Gln Glu Gln Gly Gly Ala Lys Ala

- (2) INFORMATION FOR SEQ ID NO:7:
  - (i) SEQUENCE CHARACTERISTICS:

    - (A) LENGTH: 8 amino acids
      (B) TYPE: amino acid
      (C) STRANDEDNESS: not relevant
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vii) IMMEDIATE SOURCE:
    - (B) CLONE: FLAGO peptide
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Asp Tyr Lys Asp Asp Asp Asp Lys

- (2) INFORMATION FOR SEQ ID NO:8:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 232 amino acids
    - (B) TYPE: amino acid

- (C) STRANDEDNESS: not relevant
  (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (vi) ORIGINAL SOURCE:
  (A) ORGANISM: Human
- (vii) IMMEDIATE SOURCE:
   (B) CLONE: IgG1 Fc mutein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Glu Pro Arg Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala 1 5 10 15

Pro Glu Ala Glu Gly Ala Pro Ser Val Phe Leu Phe Pro Pro Lys Pro 20 25 30

Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val 35 40 45

Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val 50 55 60

Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln 65 70 75 80

Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln 85 90 95

Asp Trp Leu Asn Gly Lys Asp Tyr Lys Cys Lys Val Ser Asn Lys Ala

Leu Pro Ala Pro Met Gln Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro

Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr

Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Arg 145 150 155 160

His Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr 165 170 175

Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr 180 185 185

Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe

Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys 210 215 220

Ser Leu Ser Leu Ser Pro Gly Lys 225 230

- (2) INFORMATION FOR SEQ ID NO:9:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 31 amino acids
    - (B) TYPE: amine acid
    - (C) STRANDEDNESS: not relevant
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: CMV (R2780 Leader)
  - (ix) FEATURE:
    - (D) OTHER INFORMATION: Metl-Arg28 is the actual leader peptide; Arg29 strengthens the furin cleavage site; nucleotides encoding Thr30 and Ser31 add a Spel site.
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:
  - Het Ala Arg Arg Leu Trp Ile Leu Ser Leu Leu Ala Val Thr Leu Thr 1 5 10  $\cdot$  15
  - Val Ala Leu Ala Ala Pro Ser Gln Lys Ser Lys Arg Arg Thr Ser 20 25 30
- (2) INFORMATION FOR SEQ ID NO:10:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 1630 base pairs

    - (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Mus musculus
  - (vii) IMMEDIATE SOURCE:

    - (A) LIBRARY: (B) CLONE: RANKL
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS
    - (B) LOCATION: 3..884
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

cc	GGC Gly 1	GTC Val	CCA Pro	CAC His	GAG Glu 5	GGT Gly	CCG Pro	CTG Leu	CAC His	CCC Pro 10	GCG Ala	CCT Pro	TCT Ser	GCA Ala	CCG Pro 15	47
GCT Ala	Pro	GCG Ala	CCG Pro	Pro 20	Pro	GCC Ala	GCC Ala	TCC Ser	CGC Arg 25	Ser	ATG Met	TTC	CTG Leu	GCC Ala 30	CTC Leu	95
CTG	GGG Gly	CTG	GGA Gly 35	Leu	GGC	CAG Gln	GTG Val	GTC Val 40	Cys	AGC Ser	ATC Ile	GCT Ala	CTG Leu 45	TTC Phe	CTG Leu	143
TAC Tyr	TTT Phe	CGA Arg 50	Ala	CAG Gln	ATG Met	gat Asp	Pro 55	Asn	AGA Arg	ATA Ile	TCA Ser	GAA Glu 60	qaA	AGC Ser	ACT Thr	191
CAC	TGC Cys 65	Pne	TAT Tyr	AGA Arg	ATC	CTG Leu 70	AGA Arg	CTC	CAT His	GAA Glu	AAC Asn 75	GCA Ala	GAT Asp	TTG Leu	CAG Gln	239
GAC Asp 80	ser	ACT Thr	CTG Leu	GAG Glu	AGT Ser 85	GAA Glu	GAC Asp	ACA Thr	CTA Leu	Pro 90	Asp	TCC Ser	TGC Cys	AGG Arg	AGG Arg 95	287
ATG Met	AAA Lys	CAA Gln	GCC Ala	TTT Phe 100	CAG Gln	GGG Gly	GCC Ala	GTG Val	CAG Gln 105	AAG Lys	GAA Glu	CTG Leu	CAA Gln	CAC His 110	ATT Ile	335
GTG Val	GGG Gly	CCA Pro	CAG Gln 115	CGC Arg	TTC Phe	TCA Ser	GGA Gly	GCT Ala 120	CCA Pro	GCT Ala	ATG Met	ATG Met	GAA Glu 125	GCC	TCA Ser	383
пр	ren	130	AWI	Ala	Gln	Arg	Gly 135	Lys	Pro	Glu	Ala	Gln 140	Pro	TIT Phe	Ala	431
CAC His	CTC Leu 145	ACC Thr	ATC Ile	AAT Asn	GCT Ala	GCC Ala 150	AGC Ser	ATC Ile	CCA Pro	TCG Ser	GGT Gly 155	TCC Ser	CAT His	AAA Lys	GTC Val	479
ACT Thr 160	CTG Leu	TCC Ser	TCT Ser	TGG Trp	TAC Tyr 165	CAC His	GAT Asp	CGA Arg	GGC Gly	TGG Trp 170	GCC Ala	AAG Lys	ATC Ile	TCT Ser	AAC Asn 175	527
Mec	inr	ren	ser	180	Gly	Lys	Leu	Arg	Val 185	Asn	Gln	Asp	Gly	TTC Phe 190	Тут	575
TAC Tyr	CTG Leu	TAC Tyr	GCC Ala 195	AAC Asn	ATT Ile	TGC Cys	TTT Phe	CGG Arg 200	CAT His	CAT His	GAA Glu	ACA Thr	TCG Ser 205	GGA Gly	AGC Ser	623
GTA Val	CCT Pro	ACA Thr 210	gac Asp	TAT Tyr	CTT Leu	CAG Gln	CTG Leu 215	ATG Met	GTG Val	TAT Tyr	GTC Val	GTT Val 220	AAA Lys	ACC Thr	AGC Ser	671
ATC Ile	AAA Lys 225	ATC Ile	CCA Pro	AGT Ser	TCT Ser	CAT His 230	AAC Asn	CTG Leu	ATG Met	AAA Lys	GGA Gly 235	GGG G1y	AGC Ser	ACG Thr	AAA Lys	719

1	AAC T Asn T 240	GG 7	rcg	GGC Gly	AAT Asn	TCT Ser 245	GAA Glu	TTC Phe	CAC His	TTT Phe	ТАТ Тут 250	TCC Ser	ATA Ile	AAT Asn	GTT Val	GGG Gly 255	767
(	GGA T	TT The I	Phe	AAG Lys	CTC Leu 260	CGA Arg	GCT Ala	GGT Gly	GAA Glu	GAA Glu 265	ATT Ile	AGC Ser	ATT Ile	CAG Gln	GTG Val 270	TCC Ser	815
i	AAC C Asn P	CT T	rcc Ser	CTG Leu 275	CTG Leu	GAT Asp	CCG Pro	GAT Asp	CAA Gln 280	GAT Asp	GCG Ala	ACG Thr	TAC Tyr	TTT Phe 285	Gly	GCT Ala	863
	MTC A Phe L	ys \						TGAG	GACT	CAT '	TTCG	rggaj	AC A	PTAG	CATG	3	914
į	ATGTC	CTAC	SA 1	GTT:	rggaj	AA C	PTCT	PAAA.	A AA'	TGGA'	TGAT	GTC	rata(	CAT	GTGT	AAGACT	974
i	ACTAA	GAGA	AC A	ATGG	CCA	G G	rgta'	rgaa.	A CT	CACA	GCCC	TCT	CTCT	IGA	GCCT	GTACAG	1034
(	STTGT	GTA1	ra 1	GTA	AAGT	CC A	ragg'	TGAT	3 TT	AGAT	TCAT	GGT	GATT	ACA	CAAC	GTTTT	1094
i	ACAAT	TTTC	3T /	LATG!	ATTI	C T	AGAA'	PTGA	A CC	AGAT"	TGGG	AGA	GGTA'	TTC	CGAT	CTTAT	1154
•	GAAAA	ACT	ra (	CACG	<b>IGAG</b> (	T A'	rgga.	AGGG	G GT	CACA	GTCT	CTG	GGTC	TAA	cccc	IGGACA	1214
•	rgrgc	CAC	rg A	GAA	CTT	GA A	ATTA	AGAGO	TA E	GCCA'	TGTC	ATT	GCAA	AGA	AATG.	ATAGTG	1274
•	TGAAG	GGT1	ra A	GTT	CTTT	rg a	ATTG	TTAC	A TT	cccc	TGGG	ACC	TGCA	TAA	AAGT	TCTTTT	1334
٠	PTTCT	YLAK	3A C	3GAG	AGAA	AA A'	rata'	TGTA'	r TT	TTAT:	ATAA	TGT	СТАА	AGT	TATA	ITTCAG	1394
(	GTGTA	ATG	rr 1	TCT	STGC	AA A	STTP	IGTA.	A AT	TATA'	TTTG	TGC	ТАТА	GTA	TTTG	аттсаа	1454
i	TATAA	TTA	AA A	ATG!	PCTC	AC TY	STTG.	ACAT	A TT	TAAT	GTTT	TAA	atgt.	ACA	GATG	TATTTA	1514
į	ACTGG	TGC	AC 2	TTG:	raat!	rc c	CCTG.	AAGG	T AC	TCGT.	AGCT	AAG	GGGG	CAG	AATA	CTGTTT	1574
(	CTGGT	GAC	CA C	CATG	PAGT.	PT A	TTTC	TTTA'	T TC	TTTT	TAAC	TTA	ATAG.	AGT	CTTC	AG	1630

#### (2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 294 amino acids
    (B) TYPE: amino acid
    (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Gly Val Pro His Glu Gly Pro Leu His Pro Ala Pro Ser Ala Pro Ala 1 5 10 15

Pro Ala Pro Pro Pro Ala Ala Ser Arg Ser Met Phe Leu Ala Leu Leu 20 25 30

Gly Leu Gly Gln Val Val Cys Ser Ile Ala Leu Phe Leu Tyr  $35 \hspace{1cm} 40 \hspace{1cm} 45$ 

Phe Arg Ala Gln Met Asp Pro Asn Arg Ile Ser Glu Asp Ser Thr His 50 60

Cys Phe Tyr Arg Ile Leu Arg Leu His Glu Asn Ala Asp Leu Gln Asp 65 70 75 80

Ser Thr Leu Glu Ser Glu Asp Thr Leu Pro Asp Ser Cys Arg Arg Met 85 90 95

Lys Gln Ala Phe Gln Gly Ala Val Gln Lys Glu Leu Gln His Ile Val 100 \$105\$

Gly Pro Gln Arg Phe Ser Gly Ala Pro Ala Met Met Glu Gly Ser Trp 115 120 125

Leu Asp Val Ala Gln Arg Gly Lys Pro Glu Ala Gln Pro Phe Ala His 130 135 140

Leu Thr Ile Asn Ala Ala Ser Ile Pro Ser Gly Ser His Lys Val Thr 145 150 155 160

Leu Ser Ser Trp Tyr His Asp Arg Gly Trp Ala Lys Ile Ser Asn Met 165 170 175

Thr Leu Ser Asn Gly Lys Leu Arg Val Asn Gln Asp Gly Phe Tyr Tyr 180 185 190

Leu Tyr Ala Asn Ile Cys Phe Arg His His Glu Thr Ser Gly Ser Val 195  $\phantom{\bigg|}200\phantom{\bigg|}$  205

Pro Thr Asp Tyr Leu Gln Leu Met Val Tyr Val Val Lys Thr Ser Ile 210 215 220

Lys Ile Pro Ser Ser His Asn Leu Met Lys Gly Gly Ser Thr Lys Asn 225 230 235 240

Trp Ser Gly Asn Ser Glu Phe His Phe Tyr Ser Ile Asn Val Gly Gly 245 250 255

Phe Phe Lys Leu Arg Ala Gly Glu Glu Ile Ser Ile Gln Val Ser Asn 260 265 270

Pro Ser Leu Leu Asp Pro Asp Gln Asp Ala Thr Tyr Phe Gly Ala Phe 275 280 285

Lys Val Gln Asp Ile Asp 290

#### (2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 954 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO

(iv)	ANTI-SENSE:	NO
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- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Homo sapiens
- (vii) IMMEDIATE SOURCE:
  - (A) LIBRARY:
  - (B) CLONE: huRANKL (full length)
- (ix) FEATURE:

  - (A) NAME/KEY: CDS (B) LOCATION: 1..951
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

ATG Met 1	CGC Arg	CGC Arg	GCC Ala	AGC Ser 5	AGA Arg	GAC Asp	TAC Tyr	ACC Thr	AAG Lys 10	TAC Tyr	CTG Leu	CGT Arg	GGC Gly	TCG Ser 15	GAG Glu	48
GAG Glu	ATG Met	GGC Gly	GGC Gly 20	GGC Gly	CCC Pro	GGA Gly	GCC Ala	CCG Pro 25	CAC His	GAG Glu	GCGC	CCC Pro	CTG Leu 30	CAC His	GCC Ala	96
CCG Pro	CCG Pro	CCG Pro	CCT Pro	GCG Ala	CCG Pro	CAC His	CAG Gln	CCC Pro	CCC Pro	GCC Ala	GCC Ala	TCC Ser	CGC Arg	TCC Ser	ATG Met	144

- TTC GTG GCC CTC CTG GGG CTG GGC CTG GGC CAG GTT GTC TGC AGC GTC
  Phe Val Ala Leu Leu Gly Leu Gly Leu Gly Gln Val Val Cys Ser Val
- GCC CTG TTC TTC TAT TTC AGA GCG CAG ATG GAT CCT AAT AGA ATA TCA Ala Leu Phe Phe Tyr Phe Arg Ala Gln Met Asp Pro Asn Arg Ile Ser 65 70 75 80 240
- GAA GAT GGC ACT CAC TGC ATT TAT AGA ATT TTG AGA CTC CAT GAA AAT Glu Asp Gly Thr His Cys Ile Tyr Arg Ile Leu Arg Leu His Glu Asn 85 90 95288
- GCA GAT TTT CAA GAC ACA ACT CTG GAG AGT CAA GAT ACA AAA TTA ATA Ala Asp Phe Gln Asp Thr Thr Leu Glu Ser Gln Asp Thr Lys Leu Ile 100 105 110
- CCT GAT TCA TGT AGG AGA ATT AAA CAG GCC TTT CAA GGA GCT GTG CAA Pro Asp Ser Cys Arg Arg Ile Lys Gln Ala Phe Gln Gly Ala Val Gln 115 120 125 384
- AAG GAA TTA CAA CAT ATC GTT GGA TCA CAG CAC ATC AGA GCA GAG AAA Lys Glu Leu Gln His Ile Val Gly Ser Gln His Ile Arg Ala Glu Lys 130 135 140
- GCG ATG GTG GAT GGC TCA TGG TTA GAT CTG GCC AAG AGG AGC AAG CTT Ala Met Val Asp Gly Ser Trp Leu Asp Leu Ala Lys Arg Ser Lys Leu 145 155 160 480
- GAA GCT CAG CCT TTT GCT CAT CTC ACT ATT AAT GCC ACC GAC ATC CCA Glu Ala Gln Pro Phe Ala His Leu Thr Ile Asn Ala Thr Asp Ile Pro 165 170 175

WU	70120	<b>74</b> 20												1	PCT/U	S <b>9</b> 7.	/237
TCT Ser	GGT Gly	TCC Ser	CAT His 180	Lys	GTG Val	AGT Ser	CTG Leu	TCC Ser 185	TCT Ser	TGG Trp	TAC Tyr	CAT His	GAT Asp 190	CGG Arg	GGT Gly		576
TGG Trp	GCC Ala	AAG Lys 195	ATC Ile	TCC Ser	AAC Asn	ATG Met	ACT Thr 200	TTT Phe	AGC Ser	AAT Asn	GGA Gly	AAA Lys 205	CTA Leu	ATA Ile	GTT Val		624
AAT Asn	CAG Gln 210	GAT Asp	GGC Gly	TTT Phe	TAT Tyr	TAC Tyr 215	CTG Leu	TAT Tyr	GCC Ala	AAC Asn	ATT Ile 220	TGC Cys	TTT Phe	CGA Arg	CAT His		672
CAT His 225	GAA Glu	ACT Thr	TCA Ser	GGA Gly	GAC Asp 230	CTA Leu	GCT Ala	ACA Thr	GAG Glu	TAT Tyr 235	CTT Leu	CAA Gln	CTA Leu	ATG Met	GTG Val 240		720
ТАС Туг	GTC Val	ACT Thr	AAA Lys	ACC Thr 245	AGC Ser	ATC Ile	AAA Lys	ATC Ile	CCA Pro 250	AGT Ser	TCT Ser	CAT His	ACC Thr	CTG Leu 255	ATG Met		768
AAA Lys	GGA Gly	GGA Gly	AGC Ser 260	ACC Thr	AAG Lys	TAT Tyr	TGG Trp	TCA Ser 265	GGG Gly	AAT Asn	TCT Ser	GAA Glu	TTC Phe 270	CAT His	TTT Phe	•	816
TAT Tyr	TCC Ser	ATA Ile 275	AAC Asn	GTT Val	GCT Gly	GGA Gly	TTT Phe 280	TTT Phe	AAG Lys	TTA Leu	CGG Arg	TCT Ser 285	GGA Gly	GAG Glu	GAA Glu		864
ATC Ile	AGC Ser 290	ATC Ile	GAG Glu	GTC Val	TCC Ser	AAC Asn 295	CCC Pro	TCC Ser	TTA Leu	CTG Leu	GAT Asp 300	CCG Pro	GAT Asp	CAG Gln	GAT Asp		912
GCA Ala 305	ACA Thr	TAC Tyr	TTT Phe	GGG Gly	GCT Ala 310	TTT Phe	AAA Lys	GTT Val	CGA Arg	GAT Asp 315	ATA Ile	GAT Asp	TGA				954

#### (2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 317 amino acids
   (B) TYPE: amino acid
   (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Met Arg Arg Ala Ser Arg Asp Tyr Thr Lys Tyr Leu Arg Gly Ser Glu  $1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15$ 

Glu Met Gly Gly Pro Gly Ala Pro His Glu Gly Pro Leu His Ala 20 25 30

Pro Pro Pro Pro Ala Pro His Gln Pro Pro Ala Ala Ser Arg Ser Met 35 40 45

Phe Val Ala Leu Leu Gly Leu Gly Leu Gly Gln Val Val Cys Ser Val 50 55 60

Ala Leu Phe Phe Tyr Phe Arg Ala Gln Met Asp Pro Asn Arg Ile Ser 65 70 75 80

Glu Asp Gly Thr His Cys Ile Tyr Arg Ile Leu Arg Leu His Glu Asn  $85 \hspace{1cm} 90 \hspace{1cm} 95$ 

Ala Asp Phe Gln Asp Thr Thr Leu Glu Ser Gln Asp Thr Lys Leu Ile 100  $$105\ \ \, 110\ \ \, 110$ 

Pro Asp Ser Cys Arg Arg Ile Lys Gln Ala Phe Gln Gly Ala Val Gln 115 120 125

Lys Glu Leu Gln His Ile Val Gly Ser Gln His Ile Arg Ala Glu Lys 130 135 140

Ala Met Val Asp Gly Ser Trp Leu Asp Leu Ala Lys Arg Ser Lys Leu 145 150 155 160

Glu Ala Gln Pro Phe Ala His Leu Thr Ile Asn Ala Thr Asp Ile Pro 165 170 175

Ser Gly Ser His Lys Val Ser Leu Ser Ser Trp Tyr His Asp Arg Gly 180  $$180\$ 

Trp Ala Lys Ile Ser Asn Met Thr Phe Ser Asn Gly Lys Leu Ile Val

Asn Gln Asp Gly Phe Tyr Tyr Leu Tyr Ala Asn Ile Cys Phe Arg His 210 215 220

His Glu Thr Ser Gly Asp Leu Ala Thr Glu Tyr Leu Gln Leu Met Val 225 230 240

Lys Gly Gly Ser Thr Lys Tyr Trp Ser Gly Asn Ser Glu Phe His Phe  $260 \hspace{1.5cm} 265 \hspace{1.5cm} 265 \hspace{1.5cm} 270 \hspace{1.5cm}$ 

Tyr Ser Ile Asn Val Gly Gly Phe Phe Lys Leu Arg Ser Gly Glu Glu 275 280 285

Ile Ser Ile Glu Val Ser Asn Pro Ser Leu Leu Asp Pro Asp Gln Asp 290 295 300

Ala Thr Tyr Phe Gly Ala Phe Lys Val Arg Asp Ile Asp 305 310 315

#### (2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1878 base pairs
  - (B) TYPE: nucleic acid (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO

	(iv	) AN	TI-S	ense	: 100											
	(vi			AL SA RGAN			ine									
	(vii	(:	A) L	ATE : IBRAI LONE	RY: I	Muri	ne P	etal	Live	er E	pith	eliu	n			
	{ix		A) N	E: AME/I OCAT:			1875									
	(xi	) SE	QUEN	CE DI	escr:	[PTI	ON: 8	SEQ :	ID N	0:14	:					
ATG Met 1	GCC Ala	CCG Pro	CGC Arg	GCC Ala 5	CGG Arg	CGG Arg	CGC Arg	CGC Arg	CAG Gln 10	CTG Leu	CCC Pro	GCG Ala	CCG Pro	CTG Leu 15	CTG Leu	48
GCG Ala	CTC Leu	TGC Cys	GTG Val 20	CTG Leu	CTC Leu	GTT Val	CCA Pro	CTG Leu 25	CAG Gln	GTG Val	ACT Thr	CTC Leu	CAG Gln 30	GTC Val	ACT Thr	96
CCT Pro	CCA Pro	TGC Cys 35	ACC Thr	CAG Gln	GAG Glu	AGG Arg	CAT His 40	TAT Tyr	GAG Glu	CAT His	CTC Leu	GGA Gly 45	CGG Arg	TGT Cys	TGC Cys	144
AGC Ser	AGA Arg 50	TGC Cys	GAA Glu	CCA Pro	GGA Gly	AAG Lys 55	TAC Tyr	CTG Leu	TCC Ser	TCT Ser	AAG Lys 60	TGC Cys	ACT Thr	CCT Pro	ACC Thr	192
TCC Ser 65	GAC Asp	AGT Ser	GTG Val	TGT Cys	CTG Leu 70	CCC Pro	TGT Cys	GGC Gly	CCC Pro	GAT Asp 75	GAG Glu	TAC Tyr	TTG Leu	GAC Asp	ACC Thr 80	240
TGG Trp	AAT Asn	GAA Glu	GAA Glu	GAT Asp 85	AAA Lys	TGC Cys	TTG Leu	CTG Leu	CAT His 90	AAA Lys	GTC Val	TGT Cys	GAT Asp	GCA Ala 95	Gly	288
AAG Lys	GCC Ala	CTG Leu	GTG Val 100	GCG Ala	GTG Val	GAT Asp	CCT Pro	GGC Gly 105	AAC Asn	CAC His	ACG Thr	GCC Ala	CCG Pro 110	CGT Arg	CGC Arg	336
TGT Cys	GCT Ala	TGC Cys 115	ACG Thr	GCT Ala	GGC Gly	TAC Tyr	CAC His 120	TGG Trp	AAC Asn	TCA Ser	GAC Asp	TGC Cys 125	GAG Glu	TGC Cys	TGC Cys	384
CGC	AGG Arg 130	AAC Asn	ACG Thr	GAG Glu	Çya Çya	GCA Ala 135	CCT Pro	GGC Gly	TTC Phe	GGA Gly	GCT Ala 140	CAG Gln	CAT His	CCC Pro	TTG Leu	432
CAG Gln 145	CTC Leu	AAC Asn	AAG Lys	GAT Asp	ACG Thr 150	GTG Val	TGC Cys	ACA Thx	CCC Pro	TGC Cys 155	CTC Leu	CTG Leu	GGC Gly	TTC Phe	TTC Phe 160	480

TCA GAT GTC TTT TCG TCC ACA GAC AAA TGC AAA CCT TGG ACC AAC TGC 528 Ser Asp Val Phe Ser Ser Thr Asp Lys Cys Lys Pro Trp Thr Asn Cys 165 170 175

ACC Thr	CTC Lev	CTI Leu	GG/ Gly 180	гy	CTA Leu	GAA Glu	GCA Ala	CAC His	Gln	GGG Gly	ACA Thr	ACG Thr	GAA Glu 190	Ser	GAT Asp	576
GTG Val	GTC Val	TGC Cys 195	Ser	TCI Ser	TCC Ser	ATG Met	ACA Thr 200	Leu	AGG Arg	AGA Arg	CCA Pro	CCC Pro 205	AAG Lys	GAG Glu	GCC Ala	624
CAG Gln	GCT Ala 210	TAT	CTG	Pro	AGT Ser	CTC Leu 215	116	GTT Val	CTG Leu	CTC Leu	CTC Leu 220	TTC Phe	ATC Ile	TCT Ser	GTG Val	672
225	VAI	vai	via	AIR	230	11e	Phe	Gly	Val	Tyr 235	Tyr	Arg	Lys	Gly	GGG Gly 240	720
2,5	nia	CTG Leu	1111	245	ASI	Leu	Trp	nak	Trp 250	Val	Asn	Asp	Ala	Суз 255	Ser	768
-	2004	AGT Ser	260	ASI	Lys	GIU	Ser	Ser 265	Gly	Asp	Arg	Суइ	Ala 270	Gly	Ser	816
*****	261	GCA Ala 275	IIII	ser	ser	GIn	Gln 280	Glu	Val	Cys	Glu	Gly 285	Ile	Leu	Leu	864
Met	290	CGG Arg	GIU	GIU	ъys	Met 295	Val	Pro	Glu	Asp	Gly 300	Ala	Gly	Val	Cys	912
305		GTG Val	суѕ	WIG	310	GIY	GIÀ	Pro	Trp	Ala 315	Glu	Val	Arg	qaA	Ser 320	960
9	1111	TTC Phe	1111	325	vaı	Ser	Glu	Val	G1u 330	Thr	Gln	Gly	Asp	Leu 335	Ser	1008
ALG	БУS	ATT Ile	340	Thr	Glu	Asp	Glu	Tyr 345	Thr	Asp	Arg	Pro	Ser 350	Gln	Pro	1056
261	1111	GGT Gly 355	ser	Leu	Leu	Leu	11e 360	Gln	Gln	Gly	Ser	<b>Lys</b> 365	Ser	Ile	Pro	1104
110	370	CAG Gln	GIU	Pro	Leu	375	Val	Gly	Glu	Asn	Asp 380	Ser	Leu	Ser	Gln	1152
385	••••	ACC Thr	GIŞ	THE	390	ser	Thr	Val	Asp	Ser 395	Glu	Gly	Cys	Asp	Phe 400	1200
ACT Thr	GAG Glu	CCT Pro	PIO	AGC Ser 405	AGA Arg	ACT Thr	GAC Asp	Ser	ATG Met 410	CCC Pro	GTG Val	TCC Ser	CCT Pro	GAA Glu 415	AAG Lys	1248

CAC	CTC Leu	ACA Thr	Lys 420	, ata	ATA Ile	GAA Glu	GGT Gly	GAC Asp 425	Ser	TGC Cys	CTC Leu	Pro	TGG Trp	Va]	GTC Val	1296
AGC Ser	Ser	AAC Asn 435	3er	ACA Thr	GAT Asp	GGC	TAC Tyr 440	Thr	GGC Gly	AGT Ser	GCG Gly	AAC Asn 445	ACT Thr	Pro	GG Gly	1344
GAG Glu	GAC Asp 450	1110	GAA Glu	CCC Pro	TTT Phe	CCA Pro 455	GGG Gly	TCC Ser	CTG Leu	AAA Lys	TGT Cys 460	Gly	CCA Pro	TTC Leu	CCC Pro	1392
CAG Gln 465	cys	GCC Ala	TAC Tyr	AGC Ser	ATG Met 470	GGC	TTT Phe	CCC Pro	AGT Ser	GAA Glu 475	GCA Ala	GCA Ala	GCC Ala	AGC Ser	ATG Met 480	.1440
GCA Ala	GAG Glu	GCG Ala	GGA Gly	GTA Val 485	CGG Arg	CCC	CAG Gln	GAC Asp	AGG Arg 490	Ala	Aab GAT	GAG Glu	AGG Arg	GGA Gly 495	GCC Ala	1488
TCA Ser	GGG Gly	TCC Ser	GGG Gly 500	AGC Ser	TCC Ser	CCC Pro	AGT Ser	GAC Asp 505	CAG Gln	CCA Pro	CCT Pro	GCC Ala	TCT Ser 510	GGG Gly	AAC	1536
GTG Val	ACT Thr	GGA Gly 515	AAC Asn	AGT Ser	AAÇ Asn	TCC Ser	ACG Thr 520	TTC Phe	ATC Ile	TCT Ser	AGC Ser	GGG Gly 525	CAG Gln	GTG Val	ATG Met	1584
AAC Asn	TTC Phe 530	AAG Lys	Gly Gly	GAC Asp	ATC Ile	ATC Ile 535	GTG Val	GTG Val	TAT Tyr	GTC Val	AGC Ser 540	CAG Gln	ACC Thr	TCG Ser	CAG Gln	1632
GAG Glu 545	GGC Gly	CCG Pro	GGT Gly	TCC Ser	GCA Ala 550	GAG Glu	CCC Pro	GAG Glu	TCG Ser	GAG Glu 555	CCC Pro	GTG Val	GGC Gly	CGC Arg	CCT Pro 560	1680
GTG Val	CAG Gln	GAG Glu	GAG Glu	ACG Thr 565	CTG Leu	GCA Ala	CAC His	AGA Arg	GAC Asp 570	TCC Ser	TTT Phe	GCG Ala	GJA GGC	ACC Thr 575	GCG Ala	1728
CCG Pro	CGC Arg	TTC Phe	CCC Pro 580	GAC Asp	GTC Val	TGT Cys	GCC Ala	ACC Thr 585	GGG Gly	GCT Ala	GGG Gly	CTG Leu	CAG Gln 590	GAG Glu	CAG Gln	1776
GGG Gly	MIG	CCC Pro 595	CGG Arg	CAG Gln	AAG Lys	GAC Asp	GGG Gly 600	ACA Thr	TCG Ser	CGG Arg	CCG Pro	GTG Val 605	CAG Gln	GAG Glu	CAG Gln	1824
GGT Gly	GGG Gly 610	GCG Ala	CAG Gln	ACT Thr	TCA Ser	CTC Leu 615	CAT	ACC Thr	CAG Gln	GGG Gly	TCC Ser 620	GGA Gly	CAA Gln	TGT Cys	GCA Ala	1872
GAA Glu 625	TGA															1878

- (2) INFORMATION FOR SEQ ID NO:15:
  - (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 625 amino acids

- (B) TYPE: amino acid
  (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:
- Met Ala Pro Arg Ala Arg Arg Arg Gln Leu Pro Ala Pro Leu Leu 1 5 10 15
- Ala Leu Cys Val Leu Leu Val Pro Leu Gln Val Thr Leu Gln Val Thr
- Pro Pro Cys Thr Gln Glu Arg His Tyr Glu His Leu Gly Arg Cys Cys 35 40 45
- Ser Arg Cys Glu Pro Gly Lys Tyr Leu Ser Ser Lys Cys Thr Pro Thr 50 55 60
- Ser Asp Ser Val Cys Leu Pro Cys Gly Pro Asp Glu Tyr Leu Asp Thr
  65 70 75 80
- Trp Asn Glu Glu Asp Lys Cys Leu Leu His Lys Val Cys Asp Ala Gly 85 90 95
- Lys Ala Leu Val Ala Val Asp Pro Gly Asn His Thr Ala Pro Arg Arg 100 105 110
- Cys Ala Cys Thr Ala Gly Tyr His Trp Asn Ser Asp Cys Glu Cys Cys 115 120 125
- Arg Arg Asn Thr Glu Cys Ala Pro Gly Phe Gly Ala Gln His Pro Leu 130 135 140
- Gln Leu Asn Lys Asp Thr Val Cys Thr Pro Cys Leu Leu Gly Phe Phe 145 150 155 160
- Ser Asp Val Phe Ser Ser Thr Asp Lys Cys Lys Pro Trp Thr Asn Cys 165 170 175
- Thr Leu Leu Gly Lys Leu Glu Ala His Gln Gly Thr Thr Glu Ser Asp 180 185 190
- Val Val Cys Ser Ser Ser Met Thr Leu Arg Arg Pro Pro Lys Glu Ala 195 200 205
- Gln Ala Tyr Leu Pro Ser Leu Ile Val Leu Leu Phe Ile Ser Val
- Val Val Val Ala Ala Ile Ile Phe Gly Val Tyr Tyr Arg Lys Gly Gly 225 230 235 240
- Lys Ala Leu Thr Ala Asn Leu Trp Asn Trp Val Asn Asp Ala Cys Ser 245 250 255
- Ser Leu Ser Gly Asn Lys Glu Ser Ser Gly Asp Arg Cys Ala Gly Ser 260 265 270
- His Ser Ala Thr Ser Ser Gln Gln Glu Val Cys Glu Gly Ile Leu Leu 275 280 285

Met Thr Arg Glu Glu Lys Met Val Pro Glu Asp Gly Ala Gly Val Cys 290 295 300

- Gly Pro Val Cys Ala Ala Gly Gly Pro Trp Ala Glu Val Arg Asp Ser 305 310 315 320
- Arg Thr Phe Thr Leu Val Ser Glu Val Glu Thr Gln Gly Asp Leu Ser 325 330 335
- Arg Lys Ile Pro Thr Glu Asp Glu Tyr Thr Asp Arg Pro Ser Gln Pro 340 345 350
- Ser Thr Gly Ser Leu Leu Leu Ile Gln Gln Gly Ser Lys Ser Ile Pro 355
- Pro Phe Gln Glu Pro Leu Glu Val Gly Glu Asn Asp Ser Leu Ser Gln 370 375 380
- Cys Phe Thr Gly Thr Glu Ser Thr Val Asp Ser Glu Gly Cys Asp Phe 385 390 395 400
- Thr Glu Pro Pro Ser Arg Thr Asp Ser Met Pro Val Ser Pro Glu Lys 405
- His Leu Thr Lys Glu Ile Glu Gly Asp Ser Cys Leu Pro Trp Val Val 420 425 430
- Ser Ser Asn Ser Thr Asp Gly Tyr Thr Gly Ser Gly Asn Thr Pro Gly 435 440 445
- Glu Asp His Glu Pro Phe Pro Gly Ser Leu Lys Cys Gly Pro Leu Pro 450 455 460
- Gln Cys Ala Tyr Ser Met Gly Phe Pro Ser Glu Ala Ala Ala Ser Met 465 470 475 480
- Ala Glu Ala Gly Val Arg Pro Gln Asp Arg Ala Asp Glu Arg Gly Ala
  485 490 495
- Ser Gly Ser Gly Ser Ser Pro Ser Asp Gln Pro Pro Ala Ser Gly Asn 500 505 510
- Val Thr Gly Asn Ser Asn Ser Thr Phe Ile Ser Ser Gly Gln Val Met 515 520 525
- Asn Phe Lys Gly Asp Ile Ile Val Val Tyr Val Ser Gln Thr Ser Gln 530 535 540
- Glu Gly Pro Gly Ser Ala Glu Pro Glu Ser Glu Pro Val Gly Arg Pro 545 550 555 560
- Val Glu Glu Glu Thr Leu Ala His Arg Asp Ser Phe Ala Gly Thr Ala 565 570 575
- Pro Arg Phe Pro Asp Val Cys Ala Thr Gly Ala Gly Leu Gln Glu Gln 580 585 590
- Gly Ala Pro Arg Gln Lys Asp Gly Thr Ser Arg Pro Val Gln Glu Gln 595 600 605

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Gly Gly Ala Gln Thr Ser Leu His Thr Gln Gly Ser Gly Gln Cys Ala 615

Glu 625

- (2) INFORMATION FOR SEQ ID NO:16:
  - (i) SEQUENCE CHARACTERISTICS:

    (A) LENGTH: 20 amino acids
    (B) TYPE: amino acid
    (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro 5 . 10

Gly Ser Thr Gly

- (2) INFORMATION FOR SEQ ID NO:17:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 5 amino acids
      (B) TYPE: amino acid

    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Asp Tyr Lys Asp Glu

- (2) INFORMATION FOR SEQ ID NO:18:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 6 amino acids
      (B) TYPE: amino acid
      (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

His His His His His His

- (2) INFORMATION FOR SEQ ID NO:19:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 33 amino acids
      (B) TYPE: amino acid

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- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

Arg Met Lys Gln Ile Glu Asp Lys Ile Glu Glu Ile Leu Ser Lys Ile 1 5 10 15

Tyr His Ile Glu Asn Glu Ile Ala Arg Ile Lys Lys Leu Ile Gly Glu  $20 \hspace{1cm} 25 \hspace{1cm} 30$ 

Arg

#### THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:

- An isolated DNA selected from the group consisting of:
- (a) a DNA encoding a protein having an amino acid sequence as set forth in SEQ ID NO: 10, wherein the protein has an amino terminus selected from the group consisting of an amino acid between amino acid 1 and amino acid 139, inclusive, and a carboxy terminus selected from the group consisting an amino acid between amino acid 290 and amino acid 294, inclusive;
- (b) a DNA encoding a protein having an amino acid sequence as set forth in SEQ ID NO: 12, wherein the protein has an amino terminus selected from the group consisting of an amino acid between amino acid 1 and amino acid 162, inclusive, and a carboxy terminus selected from the group consisting of an amino acid between amino acid 313 and amino acid 317, inclusive;
- (c) DNA molecules capable of hybridization to the DNA of (a) or (b) under stringent conditions, and which encode biologically active RANKL; and
- (d) DNA molecules encoding fragments of proteins encoded by the DNA of (a), (b) or (c).
- 2. The isolated DNA of claim 1, which encodes a RANKL polypeptide that is at least about 70% identical in amino acid sequence to the native form of RANKL as set forth in SEQ ID Nos: 10 and 12.
- The isolated DNA according to claim 1 or 2, which encodes a soluble RANKL polypeptide.
- 4. An isolated DNA encoding a soluble RANKL, selected from the group consisting of:
- (a) a DNA encoding a protein having an amino acid sequence as set forth in SEQ ID NO: 10, wherein the protein has an amino terminus selected from the group consisting of an amino acid between amino acid 48 and amino acid 139, inclusive, and a carboxy terminus selected from the group consisting an amino acid 290 and amino acid 294, inclusive;



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- (b) a DNA encoding a protein having an amino acid sequence as set forth in SEQ ID NO: 12, wherein the protein has an amino terminus selected from the group consisting of an amino acid 69 and amino acid 162, inclusive, and a carboxy terminus selected from the group consisting of an amino acid 313 and amino acid 317, inclusive;
- (c) DNA molecules capable of hybridization to the DNA of (a) or (b) under stringent conditions, and which encode biologically active RANKL; and
- (d) DNA molecules encoding fragments of proteins encoded by the DNA of (a), (b) or (c).

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•••••

5. The isolated DNA of claim 4, which further comprises a DNA encoding a polypeptide selected from the group consisting of an immunoglobulin Fc domain, an immunoglobulin Fc mutein, a FLAG<sup>TM</sup> tag, a peptide comprising at least about 6 His residues, a leucine zipper, and combinations thereof.

6. A recombinant expression vector comprising a DNA sequence according to any one of claims 1 to 6.

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7. A hot cell transformed or transfected with an expression vector according to claim 6.

: . . . .

 A process for preparing a RANKL protein, comprising culturing a host cell according to claim 7 under conditions promoting expression and recovering the RANKL.

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9. An isolated DNA selected from the group consisting of oligonucleotides of at least about 17 nucleotides in length, oligonucleotides of at least about 25 nucleotides in length, and oligonucleotides of at lest about 30 nucleotides in length, which is a fragment of the DNA of SEQ ID NO: 10 or SEQ ID NO: 12.

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An isolated RANKL polypeptide selected from the group consisting of:



- (a) a polypeptide having an amino acid sequence as set forth in SEQ ID NO: 11, wherein the polypeptide has an amino terminus selected from the group consisting of an amino acid between amino acid 1 and amino acid between amino acid 139, inclusive, and a carboxy terminus selected from the group consisting of an amino acid between amino acid 290 and 294, inclusive;
- (b) a polypeptide having an amino acid sequence as set forth in SEQ ID NO: 13, wherein the polypeptide has an amino terminus selected from the group consisting of an amino acid between amino acid 1 and amino acid 162, inclusive, and a carboxy terminus selected from the group consisting of an amino acid between amino acid 313 and 317, inclusive;

(c) a RANKL polypeptide encoded by a DNA capable of hybridization to a DNA encoding the protein of (a) or (b) under stringent conditions, and which is biologically active; and

- (d) fragments of the polypeptides of (a), (b) or (c) which are biologically active.
- 11. The protein according to claim 10, having an amino acid sequence at least about 80% identical to SEQ ID NO: 11 or SEQ ID NO: 13.
- 12. The protein according to claim 10 or 11, which is soluble RANKL.
  - 13. A soluble RANKL protein which further comprises a peptide selected from the group consisting of an immunoglobulin Fc domain, an immunoglobulin Fc mutein, a FLAG<sup>TM</sup> tag, a peptide comprising at least about 6 His residues, a leucine zipper, and combinations thereof.
  - An antibody immunoreactive with RANKL polypeptide according to claim
     10.
- 30 15. The antibody according to claim 14, which is a monoclonal antibody.



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16. A method of inducing maturation of dendritic cells (DC), comprising contacting CD1a + DC with an amount of a RANKL polypeptide sufficient to result in decreased levels of CD1b/c expression on the DC, under conditions promoting viability of the DC, and allowing the DC to mature.

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17. A method of enchanting allo-stimulatory capacity in dendritic cells (DC), comprising contacting CD1a + DC with an amount of a RANKL polypeptide sufficient to increase the allo-stimulatory capacity of the DC in a mixed lymphocyte reaction (MLR), under conditions promoting viability of the DC, and allowing the DC to present antigens to T cells.

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18. A method of promoting viability of T cells In the presence of TGFß, comprising contacting T cells that have been exposed to TGFß with an amount of a RANKL polypeptide sufficient to increase the number of T cells that remain viable in the presence of TGFß, under conditions that would promote viability of T cells in the absence of TGFß, and allowing the T cells to influence T cell tolerance.

 An isolated DNA according to claim 1, substantially as hereinbefore described.

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- 20. A recombinant expression vector according to claim 6, substantially as hereinbefore described.
- 25 21. A host cell according to claim 7, substantially as hereinbefore described.
  - 22. A polypeptide according to claim 10, substantially as hereinbefore described.
- 30 23. An antibody according to claim 14, substantially as hereinbefore described.



24. A method according to claim 16, substantially as hereinbefore described.

- 25. A method according to claim 17, substantially as hereinbefore described.
- 26. A method according to claim 18, substantially as hereinbefore described.

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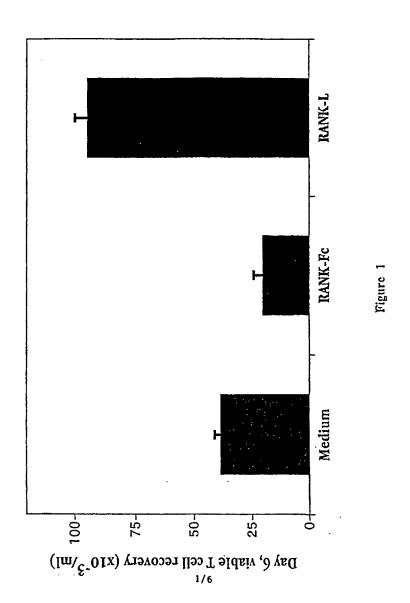
DATED: 22 June 1999

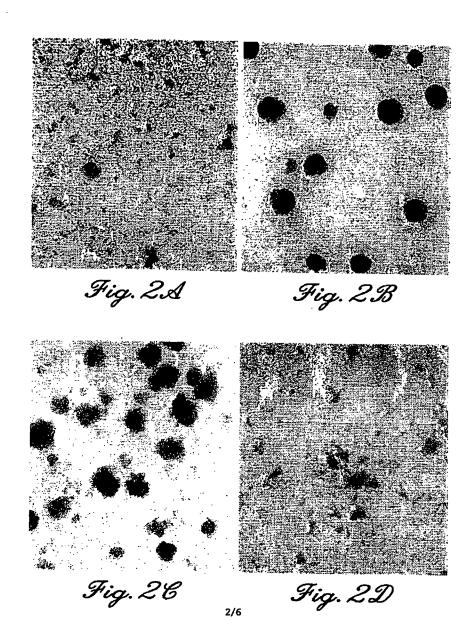
PHILLIPS ORMONDE & FITZPATRICK

Attorneys for:

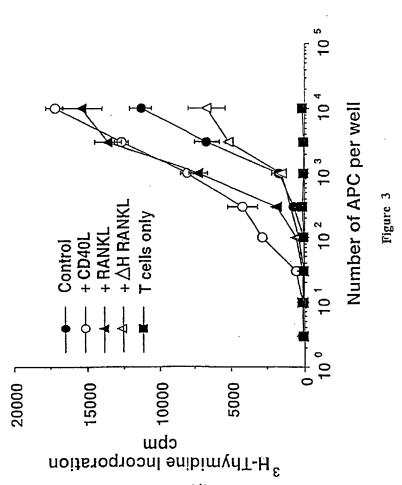
IMMUNEX CORPORATION







SUBSTITUTE SHEET (RULE 26)



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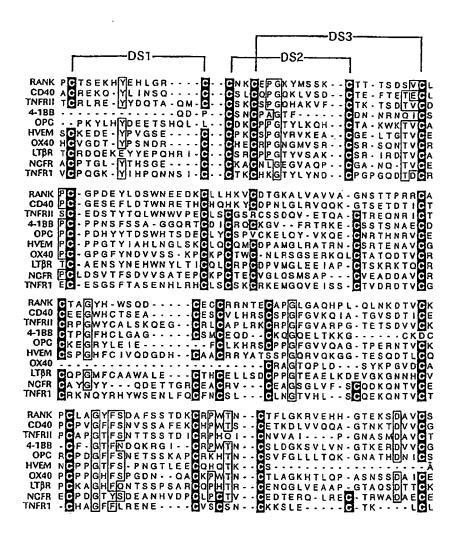


Figure 4

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Figure 5 (cont.)

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